

REC'D 17 OCT 2003

WIPO

PCT

PI 1078384

THE UNITED STATES OF AMERICA

TO ALL TO WHOM THESE PRESENTS SHALL COME:

**UNITED STATES DEPARTMENT OF COMMERCE
United States Patent and Trademark Office**

October 10, 2003

**THIS IS TO CERTIFY THAT ANNEXED HERETO IS A TRUE COPY FROM
THE RECORDS OF THE UNITED STATES PATENT AND TRADEMARK
OFFICE OF THOSE PAPERS OF THE BELOW IDENTIFIED PATENT
APPLICATION THAT MET THE REQUIREMENTS TO BE GRANTED A
FILING DATE.**

APPLICATION NUMBER: 60/426,601

FILING DATE: November 15, 2002

RELATED PCT APPLICATION NUMBER: PCT/US03/21706



**By Authority of the
COMMISSIONER OF PATENTS AND TRADEMARKS**

L. Edelen

**L. EDELEN
Certifying Officer**

**PRIORITY DOCUMENT
SUBMITTED OR TRANSMITTED IN
COMPLIANCE WITH
RULE 17.1(a) OR (b)**

11-18-02 00:26:01 11/15/02

11/15/02


11/15/02 U.S. PTO

Please type a plus sign (+) inside this box 

PTO/SB/16 (8-00)
 Approved for use through 10/31/2002 OMB 0651-0032
 U.S. Patent and Trademark Office; U.S. DEPARTMENT OF COMMERCE
 This is a request for filing a PROVISIONAL APPLICATION FOR PATENT under 37 CFR 1.53(c).

PROVISIONAL APPLICATION FOR PATENT COVER SHEET

This is a request for filing a PROVISIONAL APPLICATION FOR PATENT under 37 CFR 1.53(c).

INVENTOR(S)				
Given Name (first and middle (if any))	Family Name or Surname	Residence (City and either State or Foreign Country)		
Kevin S.	Brandt	7114 Woodrow Dr. Fort Collins, CO 80525		
<input type="checkbox"/> Additional inventors are being named on the _____ separately numbered sheets attached hereto				
TITLE OF THE INVENTION (280 characters max)				
"TICK OCTOPAMINE RECEPTOR NUCLEIC ACID MOLECULES, PROTEINS AND USES THEREOF"				
Direct all correspondence to:		CORRESPONDENCE ADDRESS		
<input checked="" type="checkbox"/> Customer Number	26949	 Place Customer Number Bar Code Here		
OR	Type Customer Number here	26949		
<input type="checkbox"/> Firm or Individual Name	PATENT TRADEMARK OFFICE			
Address				
Address				
City	State	ZIP		
Country	Telephone	Fax		
ENCLOSED APPLICATION PARTS (check all that apply)				
<input checked="" type="checkbox"/> Specification	Number of Pages	50	<input type="checkbox"/> CD(s), Number	
<input type="checkbox"/> Drawing(s)	Number of Sheets		<input checked="" type="checkbox"/> Other (specify)	12 pages Sequence Listing
<input type="checkbox"/> Application Data Sheet. See 37 CFR 1.76				
METHOD OF PAYMENT OF FILING FEES FOR THIS PROVISIONAL APPLICATION FOR PATENT (check one)				
<input checked="" type="checkbox"/>	Applicant claims small entity status. See 37 CFR 1.27.			FILING FEE AMOUNT (\$)
<input type="checkbox"/>	A check or money order is enclosed to cover the filing fees			
<input checked="" type="checkbox"/>	The Commissioner is hereby authorized to charge filing fees or credit any overpayment to Deposit Account Number			081930
<input type="checkbox"/>	Payment by credit card. Form PTO-2038 is attached.			\$80.00
The invention was made by an agency of the United States Government or under a contract with an agency of the United States Government.				
<input type="checkbox"/> No.				
<input type="checkbox"/> Yes, the name of the U.S. Government agency and the Government contract number are: _____				

Respectfully submitted,

SIGNATURE



TYPED or PRINTED NAME Timothy L. McCutcheon, Esq.

TELEPHONE

970-493-7272, ext. 4182

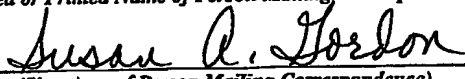
Date 11/15/02

 REGISTRATION NO. 41,184
 (if appropriate)
 Docket Number: P-FC-10

USE ONLY FOR FILING A PROVISIONAL APPLICATION FOR PATENT

This collection of information is required by 37 CFR 1.51. The information is used by the public to file (and by the PTO to process) a provisional application. Confidentiality is governed by 35 U.S.C. 122 and 37 CFR 1.14. This collection is estimated to take 8 hours to complete, including gathering, preparing, and submitting the complete provisional application to the PTO. Time will vary depending upon the individual case. Any comments on the amount of time you require to complete this form and/or suggestions for reducing this burden, should be sent to the Chief Information Officer, U.S. Patent and Trademark Office, U.S. Department of Commerce, Washington, D.C. 20231. DO NOT SEND FEES OR COMPLETED FORMS TO THIS ADDRESS. SEND TO: Box Provisional Application, Assistant Commissioner for Patents, Washington, D.C.

P18SMALL/REV05

CERTIFICATE OF MAILING BY "EXPRESS MAIL" (37 CFR 1.10) Applicant(s): Kevin S. Brandt			Docket No. P-FC-10
Serial No.	Filing Date November 15, 2002	Examiner	Group Art Unit
Invention: "TICK OCTOPAMINE RECEPTOR NUCLEIC ACID MOLECULES, PROTEINS AND USES THEREOF"			
<p>I hereby certify that the following correspondence:</p> <div style="border: 1px solid black; padding: 10px; margin: 10px 0;"> Certificate of Mailing by "Express Mail"; Original and one copy of Provisional Application for Patent Cover Sheet; 50 pages of specification, claims and abstract; 12 pages of Sequence Listing; and acknowledgment postcard </div> <p style="text-align: center;"><i>(Identify type of correspondence)</i></p> <p>is being deposited with the United States Postal Service "Express Mail Post Office to Addressee" service under 37 CFR 1.10 in an envelope addressed to: The Assistant Commissioner for Patents, Washington, D.C. 20231 on</p> <p style="text-align: center;"> <u>November 15, 2002</u> <i>(Date)</i> </p> <div style="text-align: right; margin-top: 20px;"> <p>Susan A. Gordon <i>(Typed or Printed Name of Person Mailing Correspondence)</i></p> <p> <i>(Signature of Person Mailing Correspondence)</i></p> <p>EL344802074US <i>("Express Mail" Mailing Label Number)</i></p> </div>			

TICK OCTOPAMINE RECEPTOR NUCLEIC ACID MOLECULES, PROTEINS AND USES THEREOF

FIELD OF THE INVENTION

The present invention relates to tick octopamine receptor nucleic acid molecules,
5 proteins encoded by such nucleic acid molecules, antibodies raised against such proteins, and
inhibitors of such proteins. The present invention also includes methods to obtain such
proteins, nucleic acid molecules, antibodies, and inhibitory compounds. The present
invention also includes therapeutic compositions comprising such inhibitors, as well as uses
thereof.

10 BACKGROUND OF THE INVENTION

Tick infestation of animals is a health and economic concern for pet owners. Ticks
are known to carry a variety of infectious agents including, but not limited to bacteria,
viruses, protozoan parasites, and rickettsia. As such, ticks are a problem not only when they
are on an animal but also when they are in the general environment of the animal.

15 The medical importance of tick infestation has prompted the development of reagents
capable of controlling tick infestation. Commonly encountered methods to control tick
infestation are generally focused on use of insecticides, which are often unsuccessful for one
or more of the following reasons: (1) failure of owner compliance (frequent administration is
required); (2) behavioral or physiological intolerance of the pet to the pesticide product or
20 means of administration; and (3) the emergence of tick populations resistant to the prescribed
dose of pesticide.

Octopamine receptor is a member of the biogenic amine receptor family, which also
includes receptors for dopamine, serotonin, and tyramine. Octopamine is a major
neuromodulator in insects with neurotransmitter and neurohormone functions and is a known
25 activator of adenylate cyclase, resulting in stimulation of cyclic AMP production.

Prior investigations have described certain insect biogenic amine receptors in
Drosophila melanogaster, including for example, Arakawa et al. 1990, *Neuron*, 2:343-354,

Venter et al., U.S. Patent Number 5,474,898, Saudou et al., 1990, *EMBO Journal*, 9(11):3611-3617, and Han et al., 1998, *J. Neuroscience*, 18(10):3650-3658. Unfortunately, members of the biogenic amine receptor family have proven difficult to clone due to the rarity of the sequence as cDNA and often have highly related sequences and activities, which
5 has resulted in confusion in the art with respect to the true identity of reported sequences.

Insect octopamine receptor is a known target of various insecticides, including formamidine compounds such as demethylchloridimeform (DCDM). However, no formamidine compound to date has been shown to be safe and efficacious for use for treating tick infestations on a host animal. Octopamine receptor is not present in vertebrates and
10 within insects susceptibility to insecticides targeting the octopamine receptor have been shown to vary by species. Therefore, in order to create compounds and treatments which are efficacious against ticks while minimizing toxicity to the host animal or to non-target insects, it would be a distinct advantage to have the sequence of the tick octopamine receptor. Accordingly, isolation and sequencing of tick octopamine receptor genes may be critical for
15 use in identifying specific agents for treating animals for tick infestation.

SUMMARY OF THE INVENTION

The present invention provides tick octopamine receptor proteins; nucleic acid molecules encoding tick octopamine receptor proteins; antibodies raised against such proteins (i.e., anti-tick octopamine receptor antibodies); mimetopes of such proteins or
20 antibodies; and compounds that inhibit tick octopamine receptor activity (i.e. inhibitory compounds or inhibitors).

The present invention also includes methods to obtain such proteins, mimetopes, nucleic acid molecules, antibodies and inhibitory compounds. The present invention also includes the use of proteins and antibodies to identify such inhibitory compounds as well as
25 assay kits to identify such inhibitory compounds. Also included in the present invention are therapeutic compositions comprising proteins, mimetopes, nucleic acid molecules, antibodies and inhibitory compounds of the present invention including therapeutic compounds derived

from a protein of the present invention that inhibit the activity of tick octopamine receptor proteins.

One embodiment of the present invention is an isolated tick octopamine receptor nucleic acid molecule of at least 50 nucleotides that hybridizes with a nucleic acid sequence
5 having SEQ ID NO:3, SEQ ID NO:6, SEQ ID NO:9 and SEQ ID NO:12, under conditions that allow less than or equal to 5% base pair mismatch, wherein such nucleic acid molecule encodes a protein that binds octopamine, or a nucleic acid molecule having a sequence fully complementary to such a nucleic acid molecule. Another embodiment of the present invention is an isolated tick octopamine receptor nucleic acid molecule having a nucleic acid
10 sequence that is at least 95% identical to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, and SEQ ID NO:12 or a fragment thereof having at least 50 nucleotides that encodes a protein that binds octopamine. Another embodiment of the present invention is an isolated tick octopamine receptor nucleic acid molecule that encodes a protein that is at least 95% identical to SEQ ID NO:2, SEQ ID
15 NO:5, SEQ ID NO:8 and SEQ ID NO:11, or a fragment thereof having at least 50 nucleotides that encodes a protein that binds octopamine, or a nucleic acid sequence fully complementary to such a nucleic acid sequence.

The present invention also relates to recombinant molecules, recombinant viruses and recombinant cells that include a nucleic acid molecule of the present invention. Also
20 included are methods to produce such nucleic acid molecules, recombinant molecules, recombinant viruses and recombinant cells. Also included are methods to produce a protein of the present invention.

Another embodiment of the present invention includes an isolated tick octopamine receptor protein that is at least 95% identical to an amino acid sequence selected from the
25 group consisting of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11, wherein such protein binds to octopamine.

Another embodiment of the present invention includes an isolated tick octopamine receptor protein encoded by a nucleic acid molecule that hybridizes with a nucleic acid sequence having SEQ ID NO:3, SEQ ID NO:6, SEQ ID NO:9 and SEQ ID NO:12, under conditions that allow less than or equal to 5% base pair mismatch.

5 Another embodiment of the present invention includes a method to detect an inhibitor of tick octopamine receptor activity, said method comprising (a) contacting an isolated tick octopamine receptor protein of the present invention, with a putative inhibitory compound under conditions in which, in the absence of said compound, said protein has tick octopamine receptor protein activity, and (b) determining if said putative inhibitory compound inhibits
10 tick octopamine receptor protein activity.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides for tick octopamine receptor nucleic acid molecules, proteins encoded by such nucleic acid molecules, antibodies raised against such proteins, and inhibitors of such proteins. As used herein, tick octopamine receptor nucleic acid molecules
15 and proteins encoded by such nucleic acid molecules are also referred to as octopamine receptor nucleic acid molecules and proteins of the present invention, respectively. Tick octopamine receptor nucleic acid molecules and proteins of the present invention can be isolated from a tick or prepared recombinantly or synthetically. Tick octopamine receptor nucleic acid molecules of the present invention can be RNA or DNA, or modified forms
20 thereof, and can be double-stranded or single-stranded; examples of nucleic acid molecules include, but are not limited to, complementary DNA (cDNA) molecules, genomic DNA molecules, synthetic DNA molecules, DNA molecules which are specific tags for messenger RNA, and corresponding mRNA molecules. As such, a tick nucleic acid molecule of the present invention is not intended to refer to an entire chromosome within which such a
25 nucleic acid molecule is contained, however, a tick octopamine receptor nucleic acid molecule of the present invention may include all regions such as regulatory regions that control production of tick octopamine receptor proteins encoded by such a nucleic acid

molecule (such as, but not limited to, transcription, translation or post-translation control regions) as well as the coding region itself, and any introns or non-translated coding regions. As used herein, the phrase "tick octopamine receptor protein" refers to a protein encoded by a tick octopamine receptor nucleic acid molecule.

5 Tick octopamine receptor nucleic acid molecules of known length isolated from a tick, such as *Rhipicephalus sanguineus* are denoted "nRsOCR_#", for example nRsOCR₁₄₄₃, wherein "#" refers to the number of nucleotides in that molecule, and tick octopamine receptor proteins of known length are denoted "PRsOCR_#" (for example PRsOCR₄₈₀) wherein "#" refers to the number of amino acid residues in that molecule.

10 The present invention also provides for tick octopamine receptor DNA molecules that are specific tags for messenger RNA molecules. Such DNA molecules can correspond to an entire or partial sequence of a messenger RNA, and therefore, a DNA molecule corresponding to such a messenger RNA molecule (i.e. a cDNA molecule), can encode a full-length or partial-length protein. A nucleic acid molecule encoding a partial-length
15 protein can be used directly as a probe or indirectly to generate primers to identify and/or isolate a cDNA nucleic acid molecule encoding a corresponding, or structurally related, full-length protein. A cDNA encoding a partial-length octopamine receptor protein can also be used in a similar manner to identify a genomic nucleic acid molecule, such as a nucleic acid molecule that contains the complete gene including regulatory regions, exons and introns.
20 Methods for using cDNA molecules and sequences encoding partial-length tick octopamine receptor proteins to isolate nucleic acid molecules encoding full-length tick octopamine receptor proteins and corresponding cDNA molecules are described in the examples herein below.

25 The proteins and nucleic acid molecules of the present invention can be obtained from their natural source, or can be produced using, for example, recombinant nucleic acid technology or chemical synthesis. Also included in the present invention is the use of these

proteins and nucleic acid molecules as well as antibodies and inhibitory compounds thereto as therapeutic compositions, as well as in other applications, such as those disclosed below.

One embodiment of the present invention is an isolated protein that includes a tick octopamine receptor protein. It is to be noted that the term "a" or "an" entity refers to one or more of that entity; for example, a protein, a nucleic acid molecule, an antibody and a therapeutic composition refers to "one or more" or "at least one" protein, nucleic acid molecule, antibody and therapeutic composition respectively. As such, the terms "a" (or "an"), "one or more" and "at least one" can be used interchangeably herein. It is also to be noted that the terms "comprising", "including", and "having" can be used interchangeably.

According to the present invention, an isolated, or biologically pure, protein, is a protein that has been removed from its natural milieu. As such, "isolated" and "biologically pure" do not necessarily reflect the extent to which the protein has been purified. An isolated protein of the present invention can be obtained from its natural source, can be produced using recombinant DNA technology, or can be produced by chemical synthesis.

As used herein, isolated tick octopamine receptor proteins of the present invention can be full-length proteins or any homologue of such proteins. An isolated protein of the present invention, including a homologue, can be identified in a straight-forward manner by the protein's ability to elicit an immune response against a tick octopamine receptor protein or by the protein's ability to exhibit tick octopamine receptor activity, e.g. the ability to bind to octopamine. Examples of tick octopamine receptor homologue proteins include tick octopamine receptor proteins in which amino acids have been deleted (e.g., a truncated version of the protein, such as a peptide), inserted, inverted, substituted and/or derivatized (e.g., by glycosylation, phosphorylation, acetylation, myristoylation, prenylation, palmitoylation, amidation and/or addition of glycerophosphatidyl inositol) such that the homologue includes at least one epitope capable of eliciting an immune response against a tick octopamine receptor protein, and/or of binding to an antibody directed against a tick octopamine receptor protein. That is, when the homologue is administered to an animal as an

immunogen, using techniques known to those skilled in the art, the animal will produce an immune response against at least one epitope of a natural tick octopamine receptor protein. The ability of a protein to effect an immune response can be measured using techniques known to those skilled in the art. As used herein, the term "epitope" refers to the smallest portion of a protein or other antigen capable of selectively binding to the antigen binding site of an antibody or a T cell receptor. It is well accepted by those skilled in the art that the minimal size of a protein epitope is about four to six amino acids. As is appreciated by those skilled in the art, an epitope can include amino acids that naturally are contiguous to each other as well as amino acids that, due to the tertiary structure of the natural protein, are in sufficiently close proximity to form an epitope. According to the present invention, an epitope includes a portion of a protein comprising at least 4 amino acids, at least 5 amino acids, at least 6 amino acids, at least 10 amino acids, at least 15 amino acids, at least 20 amino acids, at least 25 amino acids, at least 30 amino acids, at least 35 amino acids, at least 40 amino acids or at least 50 amino acids in length.

In one embodiment of the present invention a tick octopamine receptor homologue protein has tick octopamine receptor activity, i.e. the homologue exhibits an activity similar to its natural counterpart, e.g. the ability to bind octopamine. Methods to detect and measure such activities are known to those skilled in the art.

Tick octopamine receptor homologue proteins can be the result of natural allelic variation or natural mutation. Tick octopamine receptor protein homologues of the present invention can also be produced using techniques known in the art including, but not limited to, direct modifications to the protein or modifications to the gene encoding the protein using, for example, classic or recombinant DNA techniques to effect random or targeted mutagenesis.

Tick octopamine receptor proteins of the present invention are encoded by tick octopamine receptor nucleic acid molecules. As used herein, tick octopamine receptor nucleic acid molecules include nucleic acid sequences related to natural tick octopamine

receptor genes, and, preferably, to *R. sanguineus* tick octopamine receptor genes. As used herein, tick octopamine receptor genes include all regions such as regulatory regions that control production of tick octopamine receptor proteins encoded by such genes (such as, but not limited to, transcription, translation or post-translation control regions) as well as the coding region itself, and any introns or non-translated coding regions. As used herein, a nucleic acid molecule that "includes" or "comprises" a sequence may include that sequence in one contiguous array, or may include the sequence as fragmented exons such as is often found for a tick gene. As used herein, the term "coding region" refers to a continuous linear array of nucleotides that translates into a protein. A full-length coding region is that coding region that is translated into a full-length, i.e., a complete protein as would be initially translated in its natural milieu, prior to any post-translational modifications.

One embodiment of the present invention is a *R. sanguineus* tick octopamine receptor gene that includes the nucleic acid sequence SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, and SEQ ID NO:12 either in a consecutive array or interrupted by naturally occurring introns. These nucleic acid sequences are further described herein. For example, nucleic acid sequence SEQ ID NO:10 represents the deduced sequence of the coding strand of a *R. sanguineus* cDNA denoted herein as *R. sanguineus* octopamine receptor nucleic acid molecule nRsOCR₁₄₄₃, the production of which is disclosed in the Examples. Nucleic acid molecule SEQ ID NO:10 comprises an apparently full-length coding region. The complement of SEQ ID NO:10 (represented herein by SEQ ID NO:12) refers to the nucleic acid sequence of the strand fully complementary to the strand having SEQ ID NO:10, which can easily be determined by those skilled in the art. Likewise, a nucleic acid sequence complement of any nucleic acid sequence of the present invention refers to the nucleic acid sequence of the nucleic acid strand that is fully complementary to (i.e., can form a complete double helix with) the strand for which the sequence is cited. It should be noted that since nucleic acid sequencing technology is not entirely error-free, SEQ ID NO:10 (as well as other nucleic acid and

protein sequences presented herein) represents an apparent nucleic acid sequence of the nucleic acid molecule encoding a tick octopamine receptor protein of the present invention.

Translation of SEQ ID NO:10, the coding strand of nRsOCR₁₄₄₃ yields a protein of 480 amino acids, denoted herein as PRsOCR₄₈₀, the amino acid sequence of which is
5 presented in SEQ ID NO:11, assuming an (a) initiation codon extending from nucleotide 1 to 3 of SEQ ID NO:10 and (b) a last codon extending from nucleotide 1 to 1440 of SEQ ID NO:10.

In one embodiment, a gene or other nucleic acid molecule of the present invention can be an allelic variant that includes a similar but not identical sequence to SEQ ID NO:1,
10 SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, and SEQ ID NO:12. For example, an allelic variant of a *R. sanguineus* octopamine receptor gene including SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, and SEQ ID NO:12, is a gene that occurs at essentially the same locus (or loci) in the genome as the gene including SEQ ID NO:1, SEQ ID NO:3, SEQ
15 ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, and SEQ ID NO:12, but which, due to natural variations caused by, for example, mutation or recombination, has a similar but not identical sequence. Because natural selection typically selects against alterations that affect function, allelic variants (i.e. alleles corresponding to, or of, cited nucleic acid sequences) usually encode proteins having similar activity to that of the protein
20 encoded by the gene to which they are being compared. Allelic variants of genes or nucleic acid molecules can also comprise alterations in the 5' or 3' untranslated regions of the gene (e.g., in regulatory control regions), or can involve alternative splicing of a nascent transcript, thereby bringing alternative exons into juxtaposition. Allelic variants are well known to those skilled in the art and would be expected to occur naturally within a given tick species,
25 since the genome is diploid, and sexual reproduction will result in the reassortment of alleles.

In one embodiment of the present invention, isolated tick octopamine receptor proteins are encoded by nucleic acid molecules that hybridize under stringent hybridization

conditions to genes or other nucleic acid molecules encoding tick octopamine receptor proteins, respectively. The minimal size of tick octopamine receptor proteins of the present invention is a size sufficient to be encoded by a nucleic acid molecule capable of forming a stable hybrid (i.e., hybridizing under stringent hybridization conditions) with the
5 complementary sequence of a nucleic acid molecule encoding the corresponding natural protein. The size of a nucleic acid molecule encoding such a protein is dependent on the nucleic acid composition and the percent homology between the tick octopamine receptor nucleic acid molecule and the complementary nucleic acid sequence. It can easily be understood that the extent of homology required to form a stable hybrid under stringent
10 conditions can vary depending on whether the homologous sequences are interspersed throughout a given nucleic acid molecule or are clustered (i.e., localized) in distinct regions on a given nucleic acid molecule.

The minimal size of a nucleic acid molecule capable of forming a stable hybrid with a gene encoding a tick octopamine receptor protein is at least about 12 to about 15 nucleotides
15 in length if the nucleic acid molecule is GC-rich and at least about 15 to about 17 bases in length if it is AT-rich. The minimal size of a nucleic acid molecule used to encode a tick octopamine receptor protein homologue of the present invention is from about 12 to about 18 nucleotides in length. Thus, the minimal size of tick octopamine receptor protein homologues of the present invention is from about 4 to about 6 amino acids in length. There
20 is no limit, other than a practical limit, on the maximal size of a nucleic acid molecule encoding a tick octopamine receptor protein of the present invention because a nucleic acid molecule of the present invention can include a portion of a gene or cDNA or RNA, an entire gene or cDNA or RNA, or multiple genes or cDNA or RNA. The preferred size of a protein encoded by a nucleic acid molecule of the present invention depends on whether a full-
25 length, fusion, multivalent, or functional portion of such a protein is desired.

Stringent hybridization conditions are determined based on defined physical properties of the tick octopamine receptor nucleic acid molecule to which the nucleic acid

molecule is being hybridized, and can be defined mathematically. Stringent hybridization conditions are those experimental parameters that allow an individual skilled in the art to identify significant similarities between heterologous nucleic acid molecules. These conditions are well known to those skilled in the art. See, for example, Sambrook, *et al.*, 5 1989, *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Labs Press, and Meinkoth, *et al.*, 1984, *Anal. Biochem.* 138, 267-284, each of which is incorporated by reference herein in its entirety. As explained in detail in the cited references, the determination of hybridization conditions involves the manipulation of a set of variables including the ionic strength (M, in moles/liter), the hybridization temperature (°C), the 10 concentration of nucleic acid helix destabilizing agents (such as formamide), the average length of the shortest hybrid duplex (n), and the percent G + C composition of the fragment to which an unknown nucleic acid molecule is being hybridized. For nucleic acid molecules of at least about 150 nucleotides, these variables are inserted into a standard mathematical formula to calculate the melting temperature, or T_m , of a given nucleic acid molecule. As 15 defined in the formula below, T_m is the temperature at which two complementary nucleic acid molecule strands will disassociate, assuming 100% complementarity between the two strands:

$$T_m = 81.5^\circ\text{C} + 16.6 \log M + 0.41(\%G + C) - 500/n - 0.61(\%\text{formamide}).$$

For nucleic acid molecules smaller than about 50 nucleotides, hybrid stability is defined by 20 the dissociation temperature (T_d), which is defined as the temperature at which 50% of the duplexes dissociate. For these smaller molecules, the stability at a standard ionic strength is defined by the following equation:

$$T_d = 4(G + C) + 2(A + T).$$

A temperature of 5°C below T_d is used to detect hybridization between perfectly matched 25 molecules.

Also well known to those skilled in the art is how base pair mismatch, i.e. differences between two nucleic acid molecules being compared, including non-complementarity of

bases at a given location, and gaps due to insertion or deletion of one or more bases at a given location on either of the nucleic acid molecules being compared, will affect T_m or T_d for nucleic acid molecules of different sizes. For example, T_m decreases about 1°C for each 1% of mismatched base pairs for hybrids greater than about 150 bp, and T_d decreases about 5°C for each mismatched base pair for hybrids below about 50 bp. Conditions for hybrids between about 50 and about 150 base pairs can be determined empirically and without undue experimentation using standard laboratory procedures well known to those skilled in the art. These simple procedures allow one skilled in the art to set the hybridization conditions (by altering, for example, the salt concentration, the formamide concentration or the temperature) so that only nucleic acid hybrids with greater than a specified % base pair mismatch will hybridize. Because one skilled in the art can easily determine whether a given nucleic acid molecule to be tested is less than or greater than about 50 nucleotides, and can therefore choose the appropriate formula for determining hybridization conditions, he or she can determine whether the nucleic acid molecule will hybridize with a given gene under conditions designed to allow a desired amount of base pair mismatch.

Hybridization reactions are often carried out by attaching the nucleic acid molecule to be hybridized to a solid support such as a membrane, and then hybridizing with a labeled nucleic acid molecule, typically referred to as a probe, suspended in a hybridization solution. Examples of common hybridization reaction techniques include, but are not limited to, the well-known Southern and northern blotting procedures. Typically, the actual hybridization reaction is done under non-stringent conditions, i.e., at a lower temperature and/or a higher salt concentration, and then high stringency is achieved by washing the membrane in a solution with a higher temperature and/or lower salt concentration in order to achieve the desired stringency.

For example, if the skilled artisan wished to identify a nucleic acid molecule that hybridizes under conditions that would allow less than or equal to 30% pair mismatch with a tick octopamine receptor nucleic acid molecule of about 150 bp in length or greater, the

following conditions could preferably be used. The average G + C content of tick DNA is about 30%, as calculated from known tick nucleic acid sequences. The unknown nucleic acid molecules would be attached to a support membrane, and the 150 bp probe would be labeled, e.g. with a radioactive tag. The hybridization reaction could be carried out in a solution comprising 2X SSC in the absence of nucleic acid helix destabilizing compounds, at a temperature of about 37°C (low stringency conditions). Solutions of differing concentrations of SSC can be made by one of skill in the art by diluting a stock solution of 20X SSC (175.3 gram NaCl and about 88.2 gram sodium citrate in 1 liter of water, pH 7) to obtain the desired concentration of SSC. The skilled artisan would calculate the washing conditions required to allow up to 30% base pair mismatch. For example, in a wash solution comprising 1X SSC in the absence of nucleic acid helix destabilizing compounds, the T_m of perfect hybrids would be about 76.8°C:

$$81.5^{\circ}\text{C} + 16.6 \log (.15\text{M}) + (0.41 \times 30) - (500/150) - (0.61 \times 0) = 76.8^{\circ}\text{C}.$$

Thus, to achieve hybridization with nucleic acid molecules having about 20% base pair mismatch, hybridization washes would be carried out at a temperature of less than or equal to 56.8°C. It is thus within the skill of one in the art to calculate additional hybridization temperatures based on the desired percentage base pair mismatch, formulae and G/C content disclosed herein. For example, it is appreciated by one skilled in the art that as the nucleic acid molecule to be tested for hybridization against nucleic acid molecules of the present invention having sequences specified herein becomes longer than 150 nucleotides, the T_m for a hybridization reaction allowing up to 20% base pair mismatch will not vary significantly from 56.8°C. Similarly, to achieve hybridization with nucleic acid molecules having about 10% base pair mismatch, hybridization washes would be carried out at a temperature of less than or equal to 66.8°C and to achieve hybridization with nucleic acid molecules having about 5% base pair mismatch, hybridization washes would be carried out at a temperature of less than or equal to 71.8°C.

Furthermore, it is known in the art that there are commercially available computer programs for determining the degree of similarity between two nucleic acid or protein sequences. These computer programs include various known methods to determine the percentage identity and the number and length of gaps between hybrid nucleic acid

5 molecules or proteins. Preferred methods to determine the percent identity among amino acid sequences and also among nucleic acid sequences include analysis using one or more of the commercially available computer programs designed to compare and analyze nucleic acid or amino acid sequences. These computer programs include, but are not limited to, the SeqLab® Wisconsin Package™ Version 10.0-UNIX sequence analysis software, available

10 from Genetics Computer Group, Madison, WI (hereinafter "SeqLab"); and DNAsis® sequence analysis software, version 2.0, available from Hitachi Software, San Bruno, CA (hereinafter "DNAsis"). Such software programs represent a collection of algorithms paired with a graphical user interface for using the algorithms. The DNAsis and SeqLab software, for example, employ a particular algorithm, the Needleman-Wunsch algorithm to perform

15 pair-wise comparisons between two sequences to yield a percentage identity score, see Needleman, S.B. and Wunch, C.D., 1970, *J. Mol. Biol.*, 48, 443, which is incorporated herein by reference in its entirety. Such algorithms, including the Needleman-Wunsch algorithm, are commonly used by those skilled in the nucleic acid and amino acid sequencing art to compare sequences. A preferred method to determine percent identity among amino acid

20 sequences and also among nucleic acid sequences includes using the Needleman-Wunsch algorithm, available in the SeqLab software, using the Pairwise Comparison/Gap function with the nwsgapdna.cmp scoring matrix, the gap creation penalty and the gap extension penalties set at default values, and the gap shift limits set at maximum (hereinafter referred to as "SeqLab default parameters"). An additional preferred method to determine percent

25 identity among amino acid sequences and also among nucleic acid sequences includes using the Higgins-Sharp algorithm, available in the DNAsis software (hereinafter "DNAsis"), with the gap penalty set at 5, the number of top diagonals set at 5, the fixed gap penalty set at 10,

the k-tuple set at 2, the window size set at 5, and the floating gap penalty set at 10. A particularly preferred method to determine percent identity among amino acid sequences and also among nucleic acid sequences includes using the Needleman-Wunsch algorithm available in the SeqLab software, using the SeqLab default parameters.

5 One embodiment of the present invention includes a tick octopamine receptor protein. A preferred tick octopamine receptor protein includes a protein encoded by a nucleic acid molecule that hybridizes under conditions that preferably allow less than or equal to 30% base pair mismatch, preferably under conditions that allow less than or equal to 20% base pair mismatch, preferably under conditions that allow less than or equal to 10% base pair
10 mismatch, preferably under conditions that allow less than or equal to 8% base pair mismatch, preferably under conditions that allow less than or equal to 5% base pair mismatch or preferably under conditions that allow less than or equal to 2% base pair mismatch with a nucleic acid molecule selected from the group consisting of SEQ ID NO:3, SEQ ID NO:6, SEQ ID NO:9 and SEQ ID NO:12.

15 Another embodiment of the present invention includes a tick octopamine receptor protein encoded by a nucleic acid molecule that hybridizes under conditions comprising, (a) hybridizing in a solution comprising 1X SSC in the absence of nucleic acid helix destabilizing compounds, at a temperature of 37°C and (b) washing in a solution comprising 1X SSC in the absence of nucleic acid helix destabilizing compounds, at a temperature of
20 66.8°C, to an isolated nucleic acid molecule selected from the group consisting of SEQ ID NO:3, SEQ ID NO:6, SEQ ID NO:9 and SEQ ID NO:12.

Another preferred tick octopamine receptor protein of the present invention includes a protein that is encoded by a nucleic acid molecule that is preferably at least 70%, preferably at least 80%, preferably at least 90% identical, preferably at least 92% identical, preferably at
25 least 95% identical or preferably at least 98% identical to a nucleic acid molecule having the nucleic acid sequence SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7 and SEQ ID NO:10; also preferred are fragments (i.e. portions) of such proteins encoded by nucleic acid molecules

that are at least 50 nucleotides. Percent identity as used herein is determined using the Needleman-Wunsch algorithm, available in the SeqLab software using default parameters.

Additional preferred tick octopamine receptor proteins of the present invention include proteins having the amino acid sequence SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11, and proteins comprising homologues of a protein having the amino acid sequence SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11, wherein such a homologue comprises at least one epitope that elicits an immune response against a protein having an amino acid sequence SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11. Likewise, also preferred are proteins encoded by nucleic acid molecules comprising nucleic acid sequence SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7 and SEQ ID NO:10, or by homologues thereof.

A preferred isolated tick octopamine receptor protein of the present invention is a protein encoded by at least one of the following nucleic acid molecules: nRsOCR₁₀₂, nRsOCR₄₉₉, nRsOCR₂₈₆, and nRsOCR₁₄₄₃, or allelic variants of any of these nucleic acid molecules. Also preferred is an isolated protein encoded by a nucleic acid molecule having nucleic acid sequence SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7 and SEQ ID NO:10; or a protein encoded by an allelic variant of any of these listed nucleic acid molecules.

Preferred tick octopamine receptor proteins of the present invention include proteins having amino acid sequences that are at least 70%, preferably 80%, preferably 90%, preferably 95%, preferably at least 98%, preferably at least 99%, or preferably 100% identical to amino acid sequence SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11; and proteins encoded by allelic variants of nucleic acid molecules encoding tick octopamine receptor proteins having amino acid sequences SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11.

Preferred tick octopamine receptor proteins of the present invention include proteins selected from the group consisting of (a) a protein comprising an amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ

ID NO:11; and (b) a protein comprising an at least 20 contiguous amino acid portion identical in sequence to an at least 20 contiguous amino acid portion of an amino acid sequence of (a).

In one embodiment of the present invention, *R. sanguineus* octopamine receptor proteins comprise amino acid sequence SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11 (including, but not limited to, the proteins consisting of amino acid sequence SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11, fusion proteins and multivalent proteins), and proteins encoded by allelic variants of nucleic acid molecules encoding proteins having amino acid sequence SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11.

In one embodiment, a preferred tick octopamine receptor protein comprises an amino acid sequence of at least 30 amino acids, preferably at least 35 amino acids, preferably at least 75 amino acids, preferably at least 95 amino acids, preferably at least 150 amino acids, preferably at least 200 amino acids, preferably at least 300 amino acids, preferably at least 400 amino acids, preferably at least 450 amino acids, preferably at least 475 amino acids, or preferably at least 480 amino acids. In another embodiment, preferred tick octopamine receptor proteins comprise full-length proteins, i.e., proteins encoded by full-length coding regions, or post-translationally modified proteins thereof, such as mature proteins from which initiating methionine and/or signal sequences or "pro" sequences have been removed.

Also preferred are tick octopamine receptor proteins encoded by nucleic acid molecules having nucleic acid sequences comprising at least a portion of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7 and SEQ ID NO:10, as well as allelic variants of these nucleic acid molecules. A portion of such tick octopamine receptor nucleic acid molecule is preferably at least 50 nucleotides in length.

In another embodiment, a preferred tick octopamine receptor protein of the present invention is encoded by a nucleic acid molecule comprising at least 30 nucleotides, preferably at least 50 nucleotides, preferably at least 75 nucleotides, preferably at least 100

nucleotides, preferably at least 150 nucleotides, preferably at least 250 nucleotides, preferably at least 500 nucleotides, preferably at least 750 nucleotides, preferably at least 1000 nucleotides, preferably at least 1250 nucleotides, preferably at least 1400 nucleotides or preferably at least 1440 nucleotides in length that bind octopamine. Preferred tick
5 octopamine receptor proteins of the present invention are encoded by nucleic acid molecules comprising apparently full-length tick octopamine receptor coding region, i.e., nucleic acid molecules encoding an apparently full-length tick octopamine receptor protein, or extracellular domain.

Preferred tick octopamine receptor proteins of the present invention can be used to
10 develop inhibitors that, when administered to an animal in an effective manner, are capable of protecting that animal from tick infestation. In accordance with the present invention, the ability of an inhibitor of the present invention to protect an animal from tick infestation refers to the ability of that inhibitor to, for example, treat, ameliorate and/or prevent infestation caused by ticks. In particular, the phrase "to protect an animal from tick infestation" refers to
15 reducing the potential for tick population expansion on and around the animal (i.e., reducing the tick burden). Preferably, the tick population size is decreased, optimally to an extent that the animal is no longer bothered by ticks. A host animal, as used herein, is an animal from which ticks can feed by attaching to and feeding through the skin of the animal. Ticks, and other ectoparasites, can live on a host animal for an extended period of time or can attach
20 temporarily to an animal in order to feed. At any given time, a certain percentage of a tick population can be on a host animal whereas the remainder can be in the environment of the animal. Such an environment can include not only adult ticks, but also tick eggs and/or tick larvae. The environment can be of any size such that ticks in the environment are able to jump onto and off of a host animal. For example, the environment of an animal can include
25 plants, such as crops, from which ticks infest an animal. As such, it is desirable not only to reduce the tick burden on an animal per se, but also to reduce the tick burden in the environment of the animal.

Suitable ticks to target include any tick that is essentially incapable of causing disease in an animal administered an inhibitor of the present invention. As such, ticks to target include any tick that produces a protein that can be targeted by an inhibitory compound that inhibits a tick octopamine receptor protein function, thereby resulting in the decreased ability of the parasite to cause disease in an animal. Preferred ticks to target include ticks of the following genera: *Amblyomma*, *Dermacentor*, *Ixodes* and *Rhipicephalus*, with those of the species *Amblyomma americanum*, *Amblyomma maculatum*, *Dermacentor albipictus*, *Dermacentor andersoni*, *Dermacentor variabilis*, *Ixodes scapularis* and *Rhipicephalus sanguineus* being more preferred. Such ticks are also preferred for the isolation of proteins or nucleic acid molecules of the present invention.

Another embodiment of the present invention is an isolated nucleic acid molecule comprising a tick octopamine receptor nucleic acid molecule, i.e. a nucleic acid molecule that can be isolated from a tick cDNA library. The identifying characteristics of such nucleic acid molecules are heretofore described. A nucleic acid molecule of the present invention can include an isolated natural tick octopamine receptor gene or a homologue thereof, the latter of which is described in more detail below. A nucleic acid molecule of the present invention can include one or more regulatory regions, full-length or partial coding regions, or combinations thereof. The minimal size of a nucleic acid molecule of the present invention is a size sufficient to allow the formation of a stable hybrid (i.e., hybridization under stringent hybridization conditions) with the complementary sequence of another nucleic acid molecule. As such, the minimal size of a tick octopamine receptor nucleic acid molecule of the present invention is from 12 to 18 nucleotides in length.

In accordance with the present invention, an isolated nucleic acid molecule is a nucleic acid molecule that has been removed from its natural milieu (i.e., that has been subjected to human manipulation) and can include DNA, RNA, or derivatives of either DNA or RNA. As such, "isolated" does not reflect the extent to which the nucleic acid molecule has been purified. Isolated tick octopamine receptor nucleic acid molecules of the present

invention, or homologues thereof, can be isolated from a natural source or produced using recombinant DNA technology (e.g., polymerase chain reaction (PCR) amplification or cloning) or chemical synthesis. Isolated tick octopamine receptor nucleic acid molecules, and homologues thereof, can include, for example, natural allelic variants and nucleic acid molecules modified by nucleotide insertions, deletions, substitutions, and/or inversions in a manner such that the modifications do not substantially interfere with the nucleic acid molecule's ability to encode a tick octopamine receptor protein of the present invention.

A tick octopamine receptor nucleic acid molecule homologue can be produced using a number of methods known to those skilled in the art, see, for example, Sambrook et al., *ibid.*, which is incorporated by reference herein in its entirety. For example, nucleic acid molecules can be modified using a variety of techniques including, but not limited to, classic mutagenesis and recombinant DNA techniques such as site-directed mutagenesis, chemical treatment, restriction enzyme cleavage, ligation of nucleic acid fragments, PCR amplification, synthesis of oligonucleotide mixtures and ligation of mixture groups to "build" a mixture of nucleic acid molecules, and combinations thereof. Nucleic acid molecule homologues can be selected by hybridization with tick octopamine receptor nucleic acid molecules or by screening the function of a protein encoded by the nucleic acid molecule (e.g., ability to elicit an immune response against at least one epitope of a tick octopamine receptor protein or to effect tick octopamine receptor activity).

An isolated tick octopamine receptor nucleic acid molecule of the present invention can include a nucleic acid sequence that encodes at least one tick octopamine receptor protein of the present invention respectively, examples of such proteins being disclosed herein. Although the phrase "nucleic acid molecule" primarily refers to the physical nucleic acid molecule and the phrase "nucleic acid sequence" primarily refers to the sequence of nucleotides on the nucleic acid molecule, the two phrases can be used interchangeably, especially with respect to a nucleic acid molecule, or a nucleic acid sequence, being capable of encoding a tick octopamine receptor protein.

In one embodiment of the present invention, a preferred tick octopamine receptor nucleic acid molecule includes an isolated nucleic acid molecule that hybridizes under conditions that preferably allow less than or equal to 30% base pair mismatch, preferably under conditions that allow less than or equal to 20% base pair mismatch, preferably under
5 conditions that allow less than or equal to 10% base pair mismatch, preferably under conditions that allow less than or equal to 8% base pair mismatch, preferably under conditions that allow less than or equal to 5% base pair mismatch or preferably under conditions that allow less than or equal to 2% base pair mismatch with a nucleic acid molecule selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4,
10 SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, and SEQ ID NO:12.

One embodiment of the present invention includes a tick octopamine receptor nucleic acid molecule, wherein said nucleic acid molecule hybridizes under conditions comprising, (a) hybridizing in solution comprising 1X SSC in the absence of nucleic acid helix destabilizing compounds, at a temperature of 37°C and (b) washing in a solution comprising
15 1X SSC in the absence of nucleic acid helix destabilizing compounds, at a temperature of 74.6°C, to an isolated nucleic acid molecule selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, and SEQ ID NO:12.

Additional preferred tick octopamine receptor nucleic acid molecules of the present
20 invention include nucleic acid molecules comprising a nucleic acid sequence at least 50 nucleotides in length that is preferably at least 70%, preferably at least 80%, preferably at least 90%, preferably at least 92%, preferably at least 95%, or preferably at least 98% identical to a nucleic acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7 and SEQ ID NO:10 and encodes a protein that binds
25 octopamine.

One preferred nucleic acid molecule of the present invention includes at least a portion of nucleic acid sequence SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID

NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, and SEQ ID NO:12, as well as allelic variants of nucleic acid molecules having these nucleic acid sequences and homologues of nucleic acid molecules having these nucleic acid sequences; preferably such a homologue encodes or is complementary to a nucleic acid molecule that encodes at least one epitope that
5 elicits an immune response against a protein having an amino acid sequence SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11. Such nucleic acid molecules can include nucleotides in addition to those included in the SEQ ID NOs, such as, but not limited to, a full-length gene, a full-length coding region, a nucleic acid molecule encoding a fusion protein, or a nucleic acid molecule encoding a multivalent protective compound.

10 One embodiment of the present invention is a nucleic acid molecule comprising an isolated nucleic acid molecule having a nucleic acid sequence selected from the group consisting of: (a) a nucleic acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, and SEQ ID NO:12; and (b) a nucleic acid molecule having an at least 50 contiguous
15 nucleotide portion identical in sequence to an at least 50 contiguous nucleotide portion of a nucleic acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, and SEQ ID NO:12.

In one embodiment, a tick octopamine receptor nucleic acid molecule of the present
20 invention encodes a protein having an amino acid sequence that is at least 70%, preferably at least 80%, preferably at least 90%, preferably at least 95%, preferably at least 98%, preferably at least 99%, or preferably at least 100% identical to SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11. The present invention also includes a tick octopamine receptor nucleic acid molecule encoding a protein having at least a portion of
25 SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11, as well as allelic variants of a nucleic acid molecule encoding a protein having these sequences, including nucleic acid

molecules that have been modified to accommodate codon usage properties of the cells in which such nucleic acid molecules are to be expressed.

In another embodiment, a preferred tick octopamine receptor nucleic acid molecule of the present invention comprises a nucleic acid molecule comprising at least 30 nucleotides, preferably at least 50 nucleotides, preferably at least 75 nucleotides, preferably at least 100 nucleotides, preferably at least 150 nucleotides, preferably at least 250 nucleotides, preferably at least 500 nucleotides, preferably at least 750 nucleotides, preferably at least 1000 nucleotides, preferably at least 1250 nucleotides, preferably at least 1400 nucleotides or preferably at least 1440 nucleotides in length and encodes a protein that binds octopamine.

In another embodiment, a preferred tick octopamine receptor nucleic acid molecule encodes a protein comprising at least 30 amino acids, preferably at least 35 amino acids, preferably at least 75 amino acids, preferably at least 95 amino acids, preferably at least 150 amino acids, preferably at least 200 amino acids, preferably at least 300 amino acids, preferably at least 400 amino acids, preferably at least 450 amino acids, preferably at least 475 amino acids, or preferably at least 480 amino acids.

In another embodiment, a preferred tick octopamine receptor nucleic acid molecule of the present invention comprises an apparently full-length tick octopamine receptor coding region, i.e., the preferred nucleic acid molecule encodes an apparently full-length tick octopamine receptor protein, respectively, or a post-translationally modified protein thereof.

In one embodiment, a preferred tick octopamine receptor nucleic acid molecule of the present invention encodes a mature protein or extracellular domain.

In another embodiment, a preferred tick octopamine receptor nucleic acid molecule of the present invention comprises a nucleic acid molecule comprising SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, and SEQ ID NO:12, or a fragment thereof.

A tick octopamine receptor nucleic acid molecule of the present invention preferably comprises at least 30 nucleotides, preferably at least 50 nucleotides, preferably at least 75

nucleotides, preferably at least 100 nucleotides, preferably at least 150 nucleotides, preferably at least 250 nucleotides, preferably at least 500 nucleotides, preferably at least 750 nucleotides, preferably at least 1000 nucleotides, preferably at least 1250 nucleotides, preferably at least 1400 nucleotides or preferably at least 1440 nucleotides identical in
 5 sequence to a corresponding contiguous sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, and SEQ ID NO:12.

The phrase, a nucleic acid molecule comprising at least "x" contiguous, or consecutive nucleotides identical in sequence to at least "x" contiguous, or consecutive
 10 nucleotides of a nucleic acid molecule selected from the group consisting of SEQ ID NO:"y", refers to an "x"-nucleotide in length nucleic acid molecule that is identical in sequence to an "x"-nucleotide portion of SEQ ID NO:"y", as well as to nucleic acid molecules that are longer in length than "x". The additional length may be in the form of nucleotides that extend from either the 5' or the 3' end(s) of the contiguous identical "x"-nucleotide portion.
 15 The 5' and/or 3' extensions can include one or more extensions that have no identity to a molecule of the present invention, as well as extensions that show similarity or identity to cited nucleic acids sequences or portions thereof.

Knowing the nucleic acid sequences of certain tick octopamine receptor nucleic acid molecules of the present invention allows one skilled in the art to, for example, (a) make
 20 copies of those nucleic acid molecules, (b) obtain nucleic acid molecules including at least a portion of such nucleic acid molecules (e.g., nucleic acid molecules including full-length genes, full-length coding regions, regulatory control sequences, truncated coding regions), and (c) obtain other tick octopamine receptor nucleic acid molecules. Such nucleic acid molecules can be obtained in a variety of ways including screening appropriate expression
 25 libraries with antibodies of the present invention; traditional cloning techniques using oligonucleotide probes of the present invention to screen appropriate libraries; and PCR amplification of appropriate libraries or DNA using oligonucleotide primers of the present

invention. Preferred libraries to screen or from which to amplify nucleic acid molecules include cDNA libraries as well as genomic DNA libraries. Similarly, preferred DNA sources to screen or from which to amplify nucleic acid molecules include cDNA and genomic DNA. Techniques to clone and amplify genes are disclosed, for example, in Sambrook et al., *ibid*.

5 One embodiment of the present invention includes a recombinant vector, which includes at least one isolated nucleic acid molecule of the present invention, inserted into any vector capable of delivering the nucleic acid molecule into a host cell. Such a vector contains heterologous nucleic acid sequences, that is nucleic acid sequences that are not naturally found adjacent to nucleic acid molecules of the present invention and that
10 preferably are derived from a species other than the species from which the nucleic acid molecule(s) are derived. The vector can be either RNA or DNA, either prokaryotic or eukaryotic, and typically is a virus or a plasmid. Recombinant vectors can be used in the cloning, sequencing, and/or otherwise manipulating of tick octopamine receptor nucleic acid molecules of the present invention.

15 One type of recombinant vector, referred to herein as a recombinant molecule, comprises a nucleic acid molecule of the present invention operatively linked to an expression vector. The phrase operatively linked refers to insertion of a nucleic acid molecule into an expression vector in a manner such that the molecule is able to be expressed when transformed into a host cell. As used herein, an expression vector is a DNA or RNA
20 vector that is capable of transforming a host cell and of effecting expression of a specified nucleic acid molecule. Preferably, the expression vector is also capable of replicating within the host cell. Expression vectors can be either prokaryotic or eukaryotic, and are typically viruses or plasmids. Expression vectors of the present invention include any vectors that function (i.e., direct gene expression) in recombinant cells of the present invention, including
25 in bacterial, fungal, parasite, insect, other animal, and plant cells. Preferred expression vectors of the present invention can direct gene expression in bacterial, yeast, insect and mammalian cells, and more preferably in the cell types disclosed herein.

In particular, expression vectors of the present invention contain regulatory sequences such as transcription control sequences, translation control sequences, origins of replication, and other regulatory sequences that are compatible with the recombinant cell and that control the expression of nucleic acid molecules of the present invention. In particular, recombinant

5 molecules of the present invention include transcription control sequences. Transcription control sequences are sequences that control the initiation, elongation, and termination of transcription. Particularly important transcription control sequences are those which control transcription initiation, such as promoter, enhancer, operator and repressor sequences. Suitable transcription control sequences include any transcription control sequence that can

10 function in at least one of the recombinant cells of the present invention. A variety of such transcription control sequences are known to those skilled in the art. Preferred transcription control sequences include those that function in bacterial, yeast, or insect and mammalian cells, such as, but not limited to, *tac*, *lac*, *trp*, *trc*, *oxy-pro*, *omp/lpp*, *rrnB*, bacteriophage lambda (such as lambda p_L and lambda p_R and fusions that include such promoters),

15 bacteriophage T7, T7*lac*, bacteriophage T3, bacteriophage SP6, bacteriophage SP01, metallothionein, alpha-mating factor, *Pichia* alcohol oxidase, alphavirus subgenomic promoter, antibiotic resistance gene, baculovirus, *Heliothis zea* insect virus, vaccinia virus, herpesvirus, raccoon poxvirus, other poxvirus, adenovirus, cytomegalovirus (such as immediate early promoter), simian virus 40, retrovirus, actin, retroviral long terminal repeat,

20 Rous sarcoma virus, heat shock, phosphate and nitrate transcription control sequences as well as other sequences capable of controlling gene expression in prokaryotic or eukaryotic cells. Additional suitable transcription control sequences include tissue-specific promoters and enhancers as well as lymphokine-inducible promoters (e.g., promoters inducible by interferons or interleukins). Transcription control sequences of the present invention can also

25 include naturally occurring transcription control sequences naturally associated with ticks, such as *R. sanguineus* transcription control sequences. Suitable and preferred nucleic acid molecules to include in recombinant vectors of the present invention are as disclosed herein.

Recombinant molecules of the present invention may also (a) contain secretory signals (i.e., signal segment nucleic acid sequences) to enable an expressed tick octopamine receptor protein of the present invention to be secreted from the cell that produces the protein and/or (b) contain fusion sequences which lead to the expression of nucleic acid molecules of the present invention as fusion proteins. Examples of suitable signal segments include any signal segment capable of directing the secretion of a protein of the present invention. Preferred signal segments include, but are not limited to, tissue plasminogen activator (t-PA), interferon, interleukin, growth hormone, histocompatibility and viral envelope glycoprotein signal segments. Suitable fusion segments encoded by fusion segment nucleic acids are disclosed herein. In addition, a nucleic acid molecule of the present invention can be joined to a fusion segment that directs the encoded protein to the proteosome, such as a ubiquitin fusion segment. Eukaryotic recombinant molecules may also include intervening and/or untranslated sequences surrounding and/or within the nucleic acid sequences of nucleic acid molecules of the present invention.

Another embodiment of the present invention includes a recombinant cell comprising a host cell transformed with one or more recombinant molecules of the present invention. Transformation of a nucleic acid molecule into a cell can be accomplished by any method by which a nucleic acid molecule can be inserted into the cell. Transformation techniques include, but are not limited to, transfection, electroporation, microinjection, lipofection, adsorption, and protoplast fusion. A recombinant cell may remain unicellular or may grow into a tissue, organ or a multicellular organism. It is to be noted that a cell line refers to any recombinant cell of the present invention that is not a transgenic animal. Transformed nucleic acid molecules of the present invention can remain extrachromosomal or can integrate into one or more sites within a chromosome of the transformed (i.e., recombinant) cell in such a manner that their ability to be expressed is retained. Preferred nucleic acid molecules with which to transform a cell include tick octopamine receptor nucleic acid molecules disclosed herein.

Suitable host cells to transform include any cell that can be transformed with a nucleic acid molecule of the present invention. Host cells can be either untransformed cells or cells that are already transformed with at least one nucleic acid molecule (e.g., nucleic acid molecules encoding one or more proteins of the present invention and/or other proteins useful in the production of multivalent vaccines). Host cells of the present invention either can be endogenously (i.e., naturally) capable of producing tick octopamine receptor proteins of the present invention or can be capable of producing such proteins after being transformed with at least one nucleic acid molecule of the present invention. Host cells of the present invention can be any cell capable of producing at least one protein of the present invention, and include bacterial, fungal (including yeast), parasite (including helminth, protozoa and ectoparasite), other insect, other animal and plant cells. Preferred host cells include bacterial, mycobacterial, yeast, insect and mammalian cells. More preferred host cells include *Drosophila melanogaster* S2 cells, *Salmonella*, *Escherichia*, *Bacillus*, *Caulobacter*, *Listeria*, *Saccharomyces*, *Pichia*, *Spodoptera*, *Mycobacteria*, *Trichoplusia*, BHK (baby hamster kidney) cells, MDCK cells (Madin-Darby canine kidney cell line), CRFK cells (Crandell feline kidney cell line), CV-1 cells (African monkey kidney cell line used, for example, to culture raccoon poxvirus), COS (e.g., COS-7) cells, and Vero cells. Particularly preferred host cells are *Escherichia coli*, including *E. coli* K-12 derivatives; *Salmonella typhi*; *Salmonella typhimurium*, including attenuated strains such as UK-1 χ 3987 and SR-11 χ 4072; *Caulobacter*; *Pichia*; *Spodoptera frugiperda*; *Trichoplusia ni*; BHK cells; MDCK cells; CRFK cells; CV-1 cells; COS cells; Vero cells; and non-tumorigenic mouse myoblast G8 cells (e.g., ATCC CRL 1246). Additional appropriate mammalian cell hosts include other kidney cell lines, other fibroblast cell lines (e.g., human, murine or chicken embryo fibroblast cell lines), myeloma cell lines, Chinese hamster ovary cells, mouse NIH/3T3 cells, LMTK³¹ cells and/or HeLa cells. In one embodiment, the proteins may be expressed as heterologous proteins in myeloma cell lines employing immunoglobulin promoters.

A recombinant cell is preferably produced by transforming a host cell with one or more recombinant molecules, each comprising one or more nucleic acid molecules of the present invention operatively linked to an expression vector containing one or more transcription control sequences, examples of which are disclosed herein. The phrase
5 operatively linked refers to insertion of a nucleic acid molecule into an expression vector in a manner such that the molecule is able to be expressed when transformed into a host cell.

A recombinant cell of the present invention includes any cell transformed with at least one of any nucleic acid molecule of the present invention. Suitable and preferred nucleic acid molecules as well as suitable and preferred recombinant molecules with which
10 to transfer cells are disclosed herein.

Recombinant cells of the present invention can also be co-transformed with one or more recombinant molecules including tick octopamine receptor nucleic acid molecules encoding one or more proteins of the present invention and one or more other nucleic acid molecules encoding other protective compounds, as disclosed herein (e.g., to produce
15 multivalent vaccines).

Recombinant DNA technologies can be used to improve expression of transformed nucleic acid molecules by manipulating, for example, the number of copies of the nucleic acid molecules within a host cell, the efficiency with which those nucleic acid molecules are transcribed, the efficiency with which the resultant transcripts are translated, and the
20 efficiency of post-translational modifications. Recombinant techniques useful for increasing the expression of nucleic acid molecules of the present invention include, but are not limited to, operatively linking nucleic acid molecules to high-copy number plasmids, integration of the nucleic acid molecules into one or more host cell chromosomes, addition of vector stability sequences to plasmids, substitutions or modifications of transcription control signals
25 (e.g., promoters, operators, enhancers), substitutions or modifications of translational control signals (e.g., ribosome binding sites, Shine-Dalgarno sequences), modification of nucleic acid molecules of the present invention to correspond to the codon usage of the host cell,

deletion of sequences that destabilize transcripts, and use of control signals that temporally separate recombinant cell growth from recombinant enzyme production during fermentation. The activity of an expressed recombinant protein of the present invention may be improved by fragmenting, modifying, or derivatizing nucleic acid molecules encoding such a protein.

5 Isolated tick octopamine receptor proteins of the present invention can be produced in a variety of ways, including production and recovery of natural proteins, production and recovery of recombinant proteins, and chemical synthesis of the proteins. In one embodiment, an isolated protein of the present invention is produced by culturing a cell capable of expressing the protein under conditions effective to produce the protein, and
10 recovering the protein. A preferred cell to culture is a recombinant cell of the present invention. Effective culture conditions include, but are not limited to, effective media, bioreactor, temperature, pH and oxygen conditions that permit protein production. An effective, medium refers to any medium in which a cell is cultured to produce a tick octopamine receptor protein of the present invention. Such medium typically comprises an
15 aqueous medium having assimilable carbon, nitrogen and phosphate sources, and appropriate salts, minerals, metals and other nutrients, such as vitamins. Cells of the present invention can be cultured in conventional fermentation bioreactors, shake flasks, test tubes, microtiter dishes, and petri plates. Culturing can be carried out at a temperature, pH and oxygen content appropriate for a recombinant cell. Such culturing conditions are within the expertise
20 of one of ordinary skill in the art. Examples of suitable conditions are included in the Examples section.

 Depending on the vector and host system used for production, resultant proteins of the present invention may either remain within the recombinant cell; be secreted into the fermentation medium; be secreted into a space between two cellular membranes, such as the
25 periplasmic space in *E. coli*; or be retained on the outer surface of a cell or viral membrane.

 The phrase "recovering the protein", as well as similar phrases, refers to collecting the whole fermentation medium containing the protein and need not imply additional steps of

separation or purification. Proteins of the present invention can be purified using a variety of standard protein purification techniques, such as, but not limited to, affinity chromatography, ion exchange chromatography, filtration, electrophoresis, hydrophobic interaction chromatography, gel filtration chromatography, reverse phase chromatography, concanavalin
5 A chromatography, chromatofocusing and differential solubilization. Proteins of the present invention are preferably retrieved in "substantially pure" form. As used herein, "substantially pure" refers to a purity that allows for the effective use of the protein as a therapeutic composition or diagnostic. A therapeutic composition for animals, for example, should exhibit no substantial toxicity and preferably should be capable of stimulating the production
10 of antibodies in a treated animal.

The present invention also includes isolated (i.e., removed from their natural milieu) antibodies that selectively bind to a tick octopamine receptor protein of the present invention or a mimetope thereof (e.g., anti-tick octopamine receptor antibodies). As used herein, the term "selectively binds to" a protein refers to the ability of antibodies of the present invention
15 to preferentially bind to specified proteins and mimetopes thereof of the present invention. Binding can be measured using a variety of methods standard in the art including enzyme immunoassays (e.g., ELISA), immunoblot assays, etc.; see, for example, Sambrook et al., *ibid.*, and Harlow, et al., 1988, *Antibodies, a Laboratory Manual*, Cold Spring Harbor Labs Press; Harlow et al., *ibid.*, is incorporated by reference herein in its entirety. An anti-tick
20 octopamine receptor antibody of the present invention preferably selectively binds to a tick octopamine receptor protein, respectively, in such a way as to inhibit the function of that protein.

Isolated antibodies of the present invention can include antibodies in serum, or antibodies that have been purified to varying degrees. Antibodies of the present invention
25 can be polyclonal or monoclonal, or can be functional equivalents such as antibody fragments and genetically-engineered antibodies, including single chain antibodies or chimeric antibodies that can bind to one or more epitopes.

One embodiment of the present invention is a therapeutic composition that, when administered to an animal susceptible to tick infestation, is capable of protecting that animal from tick infestation. Therapeutic compositions of the present invention include at least one of the following protective molecules: an isolated tick octopamine receptor protein; a
5 mimetope of an isolated tick octopamine receptor protein; an isolated tick octopamine receptor nucleic acid molecule; and/or a compound derived from said isolated tick octopamine receptor protein that inhibits tick octopamine receptor protein activity, an anti-tick octopamine receptor antibody, and/or a compound that inhibits tick octopamine receptor activity. A therapeutic composition of the present invention can further comprise a
10 component selected from the group of an excipient, a carrier, and/or an adjuvant; these components are described further herein. As used herein, a protective molecule or protective compound refers to a compound that, when administered to an animal in an effective manner, is able to treat, ameliorate, and/or prevent tick infestation. Preferred ticks to target are heretofore disclosed. One example of a protective molecule is a vaccine or therapy, such as,
15 but not limited to, a naked nucleic acid vaccine or therapy, a recombinant virus vaccine or therapy, a recombinant cell vaccine or therapy, and a recombinant protein vaccine or therapy. Another example of a protective molecule is a compound that inhibits tick octopamine receptor protein activity, such as an isolated antibody that selectively binds to a tick octopamine receptor protein, a substrate analog of a tick octopamine receptor protein, anti-
20 sense-, triplex formation-, ribozyme-, and/or RNA drug-based compounds, or other inorganic or organic molecules that inhibit tick octopamine receptor protein activity. Inhibiting tick octopamine receptor protein activity can refer to the ability of a compound to reduce the activity of tick octopamine receptor proteins. Inhibiting tick octopamine receptor protein activity can also refer to the ability of a compound to reduce the amount of tick octopamine
25 receptor protein in a tick.

Another embodiment of the present invention includes a method to reduce tick infestation in an animal susceptible to tick infestation. Such a method includes the step of

administering to the animal a therapeutic molecule comprising a protective compound selected from the group consisting of (a) an isolated tick octopamine receptor protein; (b) a mimetope of an isolated tick octopamine receptor protein; (c) an isolated tick octopamine receptor nucleic acid molecule; and (d) a compound derived from an isolated tick octopamine receptor protein that inhibits tick octopamine receptor protein activity.

As used herein, the term derived, or the term derived from, refers to a peptide, antibody, mimetope, nucleic acid molecule, or other compound that was obtained directly or indirectly from a tick octopamine receptor protein or nucleic acid molecule of the present invention, e.g. a part of a protein or nucleic acid molecule or produced using a protein or nucleic acid molecule of the present invention. Methods to obtain derivatives from a tick octopamine receptor molecule of the present invention are known in the art, and as such include, but are not limited to molecular modeling of tick octopamine receptor proteins to determine active sites, and predicting from these active sites smaller fragments and/or mimetopes that retain and/or mimic these active sites, thereby inhibiting tick octopamine receptor protein activity. Other inhibitors of tick octopamine receptor activity can also be obtained in a variety of ways, including but not limited to screening of peptide or small chemical compound libraries against tick octopamine receptor proteins of the present invention; and screening of polyclonal or monoclonal antibodies to find antibodies that specifically bind tick octopamine receptor proteins of the present invention.

A tick octopamine receptor protein inhibitor of the present invention (i.e. an inhibitor of a tick octopamine receptor protein) is identified by its ability to mimic, bind to, modify, or otherwise interact with, a tick octopamine receptor protein, thereby inhibiting the activity of a natural tick octopamine receptor protein. Suitable inhibitors of tick octopamine receptor protein activity are compounds that can inhibit tick octopamine receptor protein activity in at least one of a variety of ways: (a) by binding to or otherwise interacting with or otherwise modifying tick octopamine receptor protein sites; (b) by binding to or otherwise interacting with or otherwise modifying tick octopamine receptor protein active site(s); (c) by binding to

the tick octopamine receptor protein and thus reducing the availability of the tick octopamine receptor protein in solution; (d) by mimicking a tick octopamine receptor protein; and (e) by interacting with other regions of the tick octopamine receptor protein to inhibit tick octopamine receptor protein activity, for example, by allosteric interaction.

5 Tick octopamine receptor protein inhibitors can be used directly as compounds in compositions of the present invention to treat animals as long as such compounds are not harmful to host animals being treated. Preferred tick octopamine receptor protein inhibitors of the present invention include, but are not limited to, tick octopamine receptor protein substrate analogs, and other molecules that bind to a tick octopamine receptor protein (e.g., to
10 an allosteric site) in such a manner that the activity of the tick octopamine receptor protein is inhibited. A tick octopamine receptor protein substrate analog refers to a compound that interacts with (e.g., binds to, associates with, modifies) the active site of a tick octopamine receptor protein. A preferred tick octopamine receptor protein substrate analog inhibits tick octopamine receptor protein activity. Tick octopamine receptor protein substrate analogs can
15 be of any inorganic or organic composition. Tick octopamine receptor protein substrate analogs can be, but need not be, structurally similar to a tick octopamine receptor protein natural substrate as long as they can interact with the active site of that tick octopamine receptor protein. Tick octopamine receptor protein substrate analogs can be designed using computer-generated structures of tick octopamine receptor proteins of the present invention
20 or computer structures of tick octopamine receptor protein's natural substrates. Preferred sites to model include one or more of the active sites of tick octopamine receptor proteins. Substrate analogs can also be obtained by generating random samples of molecules, such as oligonucleotides, peptides, peptidomimetic compounds, or other inorganic or organic molecules, and screening such samples for their ability to interfere with interaction between
25 tick octopamine receptor proteins and their substrates, e.g. by affinity chromatography techniques. A preferred tick octopamine receptor protein substrate analog is a tick octopamine receptor protein mimetic compound, i.e., a compound that is structurally and/or

functionally similar to a natural substrate of a tick octopamine receptor protein of the present invention, particularly to the region of the substrate that interacts with the tick octopamine receptor protein active site, but that inhibits tick octopamine receptor protein activity upon interacting with the tick octopamine receptor protein active site.

5 The present invention also includes a therapeutic composition comprising at least one protective molecule of the present invention in combination with at least one additional compound protective against one or more infectious agents and/or infestation by one or more ectoparasites.

10 In one embodiment, a therapeutic composition of the present invention can be used to protect an animal from tick infestation by administering such composition to a tick in order to prevent infestation. Such administration to the tick and/or animal could be oral, or by application to the animal's body surface (e.g. topical spot-on, or spraying onto the animal), or by application to the environment (e.g., spraying). Examples of such compositions include, but are not limited to, transgenic vectors capable of producing at least one therapeutic
15 composition of the present invention. In another embodiment a tick can ingest therapeutic compositions, or products thereof, present on the surface of or in the blood of a host animal that has been administered a therapeutic composition of the present invention.

20 In accordance with the present invention, a host animal (i.e., an animal that is or is capable of being infested with ticks) is treated by administering to the animal a therapeutic composition of the present invention in such a manner that the composition itself (e.g., a tick octopamine receptor protein inhibitor, a octopamine receptor protein synthesis suppressor (i.e., a compound that decreases the production or half-life of a octopamine receptor protein in ticks), a tick octopamine receptor protein mimetope, or a anti-tick octopamine receptor
25 antibody) or a product generated by the animal in response to administration of the composition (e.g., antibodies produced in response to administration of a tick octopamine receptor protein or nucleic acid molecule, or conversion of an inactive inhibitor "prodrug" to an active tick octopamine receptor protein inhibitor) ultimately enters the tick. A host animal

is preferably treated in such a way that the compound or product thereof is present on the body surface of the animal or enters the blood stream of the animal. Ticks are then exposed to the composition or product when they feed from the animal. For example, tick octopamine receptor protein inhibitors administered to an animal are administered in such a way that the inhibitors enter the blood stream of the animal, where they can be taken up by feeding ticks or are administered to the animal topically, where they can be taken up by contact with the treated animal.

In accordance with the present invention, reducing tick octopamine receptor protein activity in a tick can lead to a number of outcomes that reduce tick burden on treated animals and their surrounding environments. Such outcomes include, but are not limited to, (a) reducing the viability of ticks that feed from the treated animal, (b) reducing the fecundity of female ticks that feed from the treated animal, (c) reducing the reproductive capacity of male ticks that feed from the treated animal, (d) reducing the viability of eggs laid by female ticks that feed from the treated animal, (e) altering the blood feeding behavior of ticks that feed from the treated animal (e.g., ticks take up less volume per feeding or feed less frequently), (f) reducing the viability of tick larvae, for example due to the feeding of larvae from feces of ticks that feed from the treated animal, (g) altering the development of tick larvae (e.g., by decreasing feeding behavior, inhibiting growth, inhibiting (e.g., slowing or blocking) molting, and/or otherwise inhibiting maturation to adults), and/or (h) altering or decreasing the ability of ticks or tick larvae to digest a blood meal.

In order to protect an animal from tick infestation, a therapeutic composition of the present invention is administered to the animal in an effective manner such that the composition is capable of protecting that animal from tick infestation. Therapeutic compositions of the present invention can be administered to animals prior to infestation in order to prevent infestation (i.e., as a preventative vaccine) and/or can be administered to animals after infestation (i.e. as a therapy).

Therapeutic compositions of the present invention can be formulated in an excipient that the animal to be treated can tolerate. Examples of such excipients include water, saline, Ringer's solution, dextrose solution, Hank's solution, and other aqueous physiologically balanced salt solutions. Nonaqueous vehicles, such as fixed oils, sesame oil, ethyl oleate, or triglycerides may also be used. Other useful formulations include suspensions containing viscosity enhancing agents, such as sodium carboxymethylcellulose, sorbitol, or dextran. Excipients can also contain minor amounts of additives, such as substances that enhance isotonicity and chemical stability. Examples of buffers include phosphate buffer, bicarbonate buffer and Tris buffer, while examples of preservatives include thimerosal, or o-cresol, formalin and benzyl alcohol. Standard formulations can either be liquid injectables or solids which can be taken up in a suitable liquid as a suspension or solution for injection. Thus, in a non-liquid formulation, the excipient can comprise dextrose, serum albumin, preservatives, etc., to which sterile water or saline can be added prior to administration.

In one embodiment of the present invention, a therapeutic composition can include an adjuvant. Adjuvants are agents that are capable of enhancing the immune response of an animal to a specific antigen. Suitable adjuvants include, but are not limited to, cytokines, chemokines, and compounds that induce the production of cytokines and chemokines (e.g., granulocyte macrophage colony stimulating factor (GM-CSF), Flt-3 ligand, granulocyte colony stimulating factor (G-CSF), macrophage colony stimulating factor (M-CSF), colony stimulating factor (CSF), erythropoietin (EPO), interleukin 2 (IL-2), interleukin-3 (IL-3), interleukin 4 (IL-4), interleukin 5 (IL-5), interleukin 6 (IL-6), interleukin 7 (IL-7), interleukin 8 (IL-8), interleukin 10 (IL-10), interleukin 12 (IL-12), interferon gamma, interferon gamma inducing factor I (IGIF), transforming growth factor beta, RANTES (regulated upon activation, normal T cell expressed and presumably secreted), macrophage inflammatory proteins (e.g., MIP-1 alpha and MIP-1 beta), and Leishmania elongation initiating factor (LEIF)); bacterial components (e.g., endotoxins, in particular superantigens, exotoxins and cell wall components); aluminum-based salts; calcium-based salts; silica; polynucleotides;

toxoids; serum proteins, viral coat proteins; block copolymer adjuvants (e.g., Hunter's Titermax™ adjuvant (Vaxcel™, Inc. Norcross, GA), Ribi adjuvants (Ribi ImmunoChem Research, Inc., Hamilton, MT); and saponins and their derivatives (e.g., Quil A (Superfos Biosector A/S, Denmark). Protein adjuvants of the present invention can be delivered in the
5 form of the protein themselves or of nucleic acid molecules encoding such proteins using the methods described herein.

In one embodiment of the present invention, a therapeutic composition can include a carrier. Carriers include compounds that increase the half-life of a therapeutic composition in the treated animal. Suitable carriers include, but are not limited to, polymeric controlled
10 release vehicles, biodegradable implants, liposomes, bacteria, viruses, other cells, oils, esters, and glycols.

One embodiment of the present invention is a controlled release formulation that is capable of slowly releasing a composition of the present invention into an animal. As used herein, a controlled release formulation comprises a composition of the present invention in a
15 controlled release vehicle. Suitable controlled release vehicles include, but are not limited to, biocompatible polymers, other polymeric matrices, capsules, microcapsules, microparticles, bolus preparations, osmotic pumps, diffusion devices, liposomes, lipospheres, and transdermal delivery systems. Other controlled release formulations of the present invention include liquids that, upon administration to an animal, form a solid or a gel *in situ*. Preferred
20 controlled release formulations are biodegradable (i.e., bioerodible).

A preferred controlled release formulation of the present invention is capable of releasing a composition of the present invention into the blood of the treated animal at a constant rate sufficient to attain therapeutic dose levels of the composition. The therapeutic composition is preferably released over a period of time ranging from about 1 to about 12
25 months. A controlled release formulation of the present invention is capable of effecting a treatment preferably for at least 1 month, more preferably for at least 3 months, even more

preferably for at least 6 months, even more preferably for at least 9 months, and even more preferably for at least 12 months.

The efficacy of a therapeutic composition of the present invention to protect an animal from tick infestation can be tested in a variety of ways including, but not limited to challenge of the treated animal with the ticks to determine whether the treated animal is resistant to infestation. Challenge studies can include direct administration of ticks to the treated animal. In one embodiment, therapeutic compositions can be tested in animal models such as mice. Such techniques are known to those skilled in the art.

As discussed herein, one therapeutic composition of the present invention includes an inhibitor of tick octopamine receptor protein activity, i.e., a compound capable of substantially interfering with the function of a tick octopamine receptor protein. An inhibitor of tick octopamine receptor protein activity, or function, can be identified using tick octopamine receptor proteins of the present invention. A preferred inhibitor of tick octopamine receptor protein function is a compound capable of substantially interfering with the function of a tick octopamine receptor protein and which does not substantially interfere with the function of host animal octopamine receptor proteins. As used herein, a compound that does not substantially inhibit or interfere with host animal octopamine receptor proteins is one that, when administered to a host animal, the host animal shows no significant adverse effects attributable to the inhibition of octopamine receptor and which, when administered to an animal in an effective manner, is capable of protecting that animal from tick infestation.

One embodiment of the present invention is a method to identify a compound capable of inhibiting tick octopamine receptor protein activity. Such a method includes the steps of (a) contacting (e.g., combining, mixing) an isolated tick octopamine receptor protein of the present invention, with a putative inhibitory compound under conditions in which, in the absence of the compound, the protein has tick octopamine receptor protein activity, and (b) determining if the putative inhibitory compound inhibits the activity. Tick octopamine receptor protein activity can be determined in a variety of ways known in the art, including

but not limited to determining the ability of tick octopamine receptor protein to bind to or otherwise interact with a substrate. Such conditions under which a tick octopamine receptor protein has tick octopamine receptor protein activity include conditions in which a tick octopamine receptor protein has a correct three-dimensionally folded structure under
5 physiologic conditions, i.e. physiologic pH, physiologic ionic concentrations, and physiologic temperatures, such as a native protein, a mature protein, a soluble protein, transfected cells or viruses. Accordingly, the correct three-dimensionally folded structure could be used to predict inhibitory compounds.

Putative inhibitory compounds to screen include antibodies (including fragments and
10 mimetopes thereof), putative substrate analogs, and other, preferably small, organic or inorganic molecules. Methods to determine tick octopamine receptor protein activity are known to those skilled in the art, see for example Han et al. 1996, Neuron 16:1127-1135.

A preferred method to identify a compound capable of inhibiting tick octopamine receptor protein activity includes contacting an isolated tick octopamine receptor protein of
15 the present invention with a putative inhibitory compound under conditions in which, in the absence of the compound, the protein has tick octopamine receptor protein activity; and determining if the putative inhibitory compound inhibits the activity.

A preferred method to identify a compound capable of inhibiting tick octopamine receptor protein activity includes contacting a recombinant cell comprising an isolated tick
20 octopamine receptor protein of the present invention with a putative inhibitory compound under conditions in which, in the absence of the compound, the protein has tick octopamine receptor protein activity; and determining if the putative inhibitory compound inhibits the activity.

Another embodiment of the present invention is an assay kit to identify an inhibitor of
25 a tick octopamine receptor protein of the present invention. This kit comprises an isolated tick octopamine receptor protein of the present invention, and a means for determining inhibition of an activity of tick octopamine receptor protein, where the means enables

detection of inhibition. Detection of inhibition of tick octopamine receptor protein identifies a putative inhibitor to be an inhibitor of a tick octopamine receptor protein. Means for determining inhibition of a tick octopamine receptor protein include, for example, an assay system that detects binding of a putative inhibitor to a tick octopamine receptor molecule, and an assay system that detects interference by a putative inhibitor of the ability of tick octopamine receptor protein to bind octopamine. Means and methods are described herein and are known to those skilled in the art.

The following examples are provided for the purposes of illustration and are not intended to limit the scope of the present invention. The following examples include a number of recombinant DNA and protein chemistry techniques known to those skilled in the art; see, for example, Sambrook et al., *ibid*.

Example 1

This example describes the preparation of tick genomic DNA and a tick head and nerve cord cDNA pool by Rapid Amplification of cDNA Ends (RACE pool).

Genomic DNA was isolated from adult ticks as follows. Four adult *Rhipicephalus sanguineus* were crushed in a mortar & pestle using DNAzol™ reagent, available from Life Technologies, Rockville, MD, and genomic DNA was recovered according to manufacturer's instructions. Resultant DNA was resuspended in 50 µL T.E. and had an estimated concentration of 50ng/µL.

A tick cDNA RACE pool was constructed as follows. Approximately 10 unfed adult *Rhipicephalus sanguineus* were ground to a homogenized powder by mortar and pestle in liquid nitrogen. Total RNA was isolated from the resulting homogenized powder using a modification of the acid-guanidinium-phenol-chloroform method described by Chomczynski et al., 1987, Anal. Biochem. 162, p. 156-159, which is incorporated herein by reference in its

entirety. The modification of the method is that solution D described by Chomczynski et al. was changed to a solution of 4 M guanidine isothiocyanate, 25 mM Sodium Citrate pH 7.0, 1.5% Sarcosyl, 0.5 M 2-mercaptoethanol. Spectrophotometer and ethidium bromide stained denaturing gel analyses indicated that the yield of total RNA was approximately 43 µg.

5 Approximately 10 µg of total RNA was used as template to construct a RACE pool using a Marathon cDNA Amplification Kit, available from Clontech Laboratories, Inc., Palo Alto, CA, according to the manufacturer's instructions.

Example 2

This example describes the cloning and sequencing of tick octopamine receptor
10 nucleic acid molecules.

PCR primers whose sequence was obtained from Van Poyer et al., Insect Biochemistry and Molecular Biology, 31 (2001) 333-338 were used to amplify an approximately 100 nucleotide product from genomic DNA of adult, unfed ticks prepared as described above. Forward primer 5' GCCATCATYG TGGGCRKSTT CATCKTBTGC
15 TGG 3', designated SEQ ID NO:13, was used in combination with reverse primer 5' GATCATSGGR TTWAYGGCSG AGTTGCAGTA GCC 3', designated SEQ ID NO:14, in a PCR reaction using 2µL of genomic DNA, 2.5 units (U) AmpliTaq polymerase, available from PE Biosystems, Foster City, CA, 0.2 mM dNTP's and 1.0 µM primers, in a total reaction volume of 50 µL. The following amplification conditions were used: (1) one cycle
20 of 95°C for 1 minute, (2) five cycles of 94°C for 10 seconds, 60°C for 30 seconds, and 72°C for 30 seconds, (3) ten cycles of 94°C for 10 seconds, 57°C for 30 seconds, and 72°C for 30 seconds, (4) twenty-five cycles of 94°C for 10 seconds, 54°C for 30 seconds, and 72°C for 45 seconds. The resulting product, referred to as nRsOCR₁₀₂, was excised from an agarose gel

and T/A cloned using the Topo TA cloning kitTM from Invitrogen and sequenced to reveal a 102 nucleotide product, having a coding strand designated SEQ ID NO:1 and a complementary strand designated SEQ ID NO:3.

First and second PCR reactions were performed on the tick cDNA library prepared as described in Example 1 under the following reaction conditions: 2.5 U AdvantageTM polymerase per reaction, available from Clontech, Palo Alto, CA, 0.2 mM dNTP's, and 0.5 μ M primers, in a total reaction volume of 50 μ L, were used under the following cycling conditions: (1) one cycle of 94°C for 30 seconds, (2) five cycles of 94°C for 5 seconds, 64°C for 4 minutes, (3) five cycles of 94°C for 5 seconds, 66°C for 4 minutes, (4) five cycles of 94°C for 5 seconds, 68°C for 4 minutes, and (5) one cycle of 72°C for three minutes. In the first PCR reaction, a forward primer having the sequence 5' GCTGGCTGCC ATTCTTCACC GTG 3', designated SEQ ID NO:15 was used in combination with a primer corresponding to sequence within the adapter flanking the termini of all fragments in the cDNA RACE pool, having the sequence 5' CCATCCTAAT ACGACTCACT ATAGGGC 3', designated SEQ ID NO:16. Five μ L of a 1/250 dilution of tick cDNA RACE pool was used as template. One μ L of the resulting reaction product was used in a second PCR reaction using SEQ ID NO:16 as the reverse primer in combination with a forward primer having the sequence 5' GGTGCGTGCA TTCTGCGAGC ACTG 3', designated SEQ ID NO:17. The resulting PCR product was excised from an agarose gel and T/A cloned using the TOPO T/ATM cloning kit, available from Invitrogen, Carlsbad, CA. The purified product, denoted nRsOCR₄₉₉, was sequenced and shown to contain 499 base pairs, having a coding strand designated SEQ ID NO:4 and a complementary strand designated SEQ ID NO:6. Sequence analysis of SEQ ID NO:4 indicates that nRsOCR₄₉₉ encodes a protein denoted PRsOCR₉₂,

having a sequence represented by SEQ ID NO:5, assuming an open reading frame extending from nucleotide 3 through nucleotide 278 of SEQ ID NO:4. Sequence analysis further demonstrated that SEQ ID NO:5 represents the C-terminus of a tick octopamine receptor.

To isolate the 5' portion of a tick octopamine receptor, first and second PCR reactions were performed on the tick cDNA library prepared as described in Example 1 under the following reaction conditions: 2.5 U AdvantageTM polymerase per reaction, 0.2 mM dNTP's, and 0.5 μ M primers, in a total reaction volume of 50 μ L, were used under the following cycling conditions: (1) one cycle of 95°C for 1 minute, (2) five cycles of 94°C for 10 seconds, 66°C for 30 seconds, 72°C for 1 minute, (3) thirty cycles of 94°C for 10 seconds, 63°C for 20 seconds, 72°C for two minutes.

In the first PCR reaction, a forward primer SEQ ID NO:16 was used in combination with a reverse primer having nucleotide sequence 5' AGAAGACCGA GAACAGCAGG TTGG 3', designated SEQ ID NO:18. Three μ L of a 1/250 dilution of tick cDNA RACE pool was used as template. One μ L of the resulting reaction product was used as the template in a second PCR reaction using SEQ ID NO:16 as the reverse primer in combination with a forward primer having the sequence 5' TGGCACCAGG TGTGGCCGAA GAGCCACAC 3', designated SEQ ID NO:19. The resulting PCR product was excised from an agarose gel and T/A cloned using a TOPO T/ATM cloning kit. The purified product, denoted nRsOCR₂₈₆, was sequenced and shown to contain 286 base pairs, having a coding strand designated SEQ ID NO:7 and a complementary strand designated SEQ ID NO:9. Sequence analysis of SEQ ID NO:7 indicates that nRsOCR₂₈₆ encodes a protein denoted PRsOCR₉₅, having a sequence represented by SEQ ID NO:8, assuming an open reading frame extending from nucleotide 1 through nucleotide 285 of SEQ ID NO:7.

Sequence analysis further demonstrated that SEQ ID NO:8 represents the N-terminus of a tick octopamine receptor.

Sequence information from SEQ ID NO:3 and SEQ ID NO:6 was used to design primers to amplify one contiguous piece of DNA encoding the entire open reading frame of the nucleic acid molecule encoding a tick octopamine receptor and a PCR reaction was conducted as follows. A forward primer having nucleotide sequence 5' ATGAACGAGA CGTGCCTGTC CCGC 3', designated SEQ ID NO:20 was used in conjunction with a reverse primer having nucleotide sequence, 5' CTAGGGCGAC GCGGCGTTGT CCGG 3', designated SEQ ID NO:21. The following reaction conditions were used: 2.5 U Advantage™ polymerase per reaction, five µL of a 1/250 dilution of tick cDNA RACE pool as template, 0.2 mM dNTP, and 0.5 µM primers, in a total reaction volume of 50 µL. The following cycling conditions were used: (1) one cycle of 94°C for 5 minutes, (2) five cycles of 94°C for 5 seconds, 72°C for 2.5 minutes, (3) 5 cycles of 94°C 5 seconds, 70°C 2.5 minutes, (4) thirty-five cycles of 94°C for 5 seconds, 68°C for 2.5 minutes, and (5) one cycle of 72°C for 7 minutes. The resulting approximately 1500 nucleotide PCR product was excised from an agarose gel, purified using a Qiaquick™ kit, available from Qiagen, Chatsworth, CA, and T/A cloned using a TOPO T/A cloning kit. The purified product, referred to as nRsOCR₁₄₄₃ was sequenced and shown to contain 1443 base pairs, having a coding strand designated SEQ ID NO:10 and a complementary strand designated SEQ ID NO:12. Sequence analysis of SEQ ID NO:10 indicates that nRsOCR₁₄₄₃ encodes a protein denoted PRsOCR₄₈₀, having a sequence represented by SEQ ID NO:11, assuming an open reading frame extending from nucleotide 1 through nucleotide 1440 of SEQ ID NO:10.

Sequence analysis further demonstrated that SEQ ID NO:11 represents an open reading frame encoding a full-length tick octopamine receptor.

Comparison of nucleic acid SEQ ID NO:10 with nucleic acid sequences reported in GenBank indicates that SEQ ID NO:10 shared the most similarity, i.e. about 57% with a *Balanus amphitrite* gene for G protein-coupled receptor (GenBank accession number D78587). Comparison of amino acid sequence SEQ ID NO:11 with amino acid sequences reported in GenBank indicates that SEQ ID NO:11 showed the most similarity, i.e. about 52%, with an *Anopheles gambiae* str. PEST (genome seq conceptual translation) and a second highest similarity to a *Balanus amphitrite* G-protein coupled receptor (GenBank accession number BAA11424).

While various embodiments of the present invention have been described in detail, it is apparent that modifications and adaptations of those embodiments will occur to those skilled in the art. It is to be expressly understood, however, that such modifications and adaptations are within the scope of the present invention, as set forth in the following claims:

What is claimed is:

1. An isolated nucleic acid molecule selected from the group consisting of:
 - (a) a nucleic acid molecule at least 50 nucleotides in length that hybridizes with a
5 nucleic acid molecule having a nucleic acid sequence selected from the group consisting of
SEQ ID NO:3, SEQ ID NO:6, SEQ ID NO:9 and SEQ ID NO:12, under conditions
comprising: (1) hybridizing in a solution comprising 2X SSC in the absence of nucleic acid
helix destabilizing compounds, at a temperature of 37°C; and (2) washing in a solution
comprising 1X SSC in the absence of helix destabilizing compounds, at a temperature of
10 66.8°C; wherein said at least 50 nucleotide nucleic acid molecule encodes a protein that binds
to octopamine; and
 - (b) a nucleic acid molecule fully complementary to the nucleic acid molecule of (a).
2. The nucleic acid molecule of Claim 1, wherein said nucleic acid molecule comprises
a nucleic acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3,
15 SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, and SEQ ID
NO:12.
3. The nucleic acid molecule of Claim 1, wherein said nucleic acid molecule encodes a
protein comprising an amino acid sequence selected from the group consisting of SEQ ID
NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11.
- 20 4. A recombinant molecule comprising a nucleic acid molecule as set forth in Claim 1
operatively linked to a transcription control sequence.
5. A recombinant virus comprising a nucleic acid molecule as set forth in Claim 1.
6. A recombinant cell comprising a nucleic acid molecule as set forth in Claim 1.

7. A method to produce a protein encoded by an isolated nucleic acid molecule of Claim 1, said method comprising culturing a cell transformed with a nucleic acid molecule encoding said protein.

8. An isolated nucleic acid molecule selected from the group consisting of:

5 (a) a nucleic acid molecule having a nucleic acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7 AND SEQ ID NO:10, and variants thereof at least 95% identical to a nucleic acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7 AND SEQ ID NO:10, wherein said nucleic acid molecule encodes a protein that binds to octopamine;

10 (b) a nucleic acid molecule comprising a fragment of a nucleic acid molecule of (a), wherein said fragment is at least 50 nucleotides in length; and

(c) a nucleic acid molecule fully complementary to a nucleic acid molecule of (a) or (b).

9. The nucleic acid molecule of Claim 8, wherein said nucleic acid molecule comprises
15 a nucleic acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, and SEQ ID NO:12.

10. The nucleic acid molecule of Claim 8, wherein said nucleic acid molecule encodes a protein comprising an amino acid sequence selected from the group consisting of SEQ ID
20 NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11.

11. A recombinant molecule comprising a nucleic acid molecule as set forth in Claim 8 operatively linked to a transcription control sequence.

12. A recombinant virus comprising a nucleic acid molecule as set forth in Claim 8.

13. A recombinant cell comprising a nucleic acid molecule as set forth in Claim 8.

14. A method to produce a protein encoded by an isolated nucleic acid molecule of Claim 8, said method comprising culturing a cell transformed with a nucleic acid molecule encoding said protein.
15. An isolated protein comprising an amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11, and variants thereof that are at least 95% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11, wherein said protein variant binds to octopamine.
16. The protein of Claim 15, wherein said protein is encoded by a nucleic acid molecule having a nucleic acid sequence selected from the group consisting of: SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7 AND SEQ ID NO:10.
17. The protein of Claim 15, wherein said protein comprises an amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11.
18. An isolated antibody that selectively binds to a protein as set forth in Claim 15.
19. A method to detect an inhibitor of tick octopamine receptor activity, said method comprising (a) contacting an isolated tick octopamine receptor protein of Claim 15, with a putative inhibitory compound under conditions in which, in the absence of said compound, said protein has tick octopamine receptor protein activity, and (b) determining if said putative inhibitory compound inhibits tick octopamine receptor protein activity.
20. The method of Claim 19, wherein said protein comprises an amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8 and SEQ ID NO:11.

ABSTRACT

The present invention relates to tick octopamine receptor nucleic acid molecules; to tick octopamine receptor proteins encoded by such nucleic acid molecules; to antibodies raised against such proteins; and to compounds that inhibit the activity of such proteins. The present invention also includes methods to obtain such proteins, nucleic acid molecules, antibodies, and inhibitory compounds. The present invention also includes inhibitory compounds, particularly those that specifically inhibit tick octopamine receptor activity, as well as the use of such compounds to treat animals.

10

**VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL ENTITY STATUS
(37 CFR 1.9(f) and 1.27(c)) - SMALL BUSINESS CONCERN**

I hereby declare that I am an official empowered to act on behalf of Heska Corporation of 1613 Prospect Parkway, Fort Collins, Colorado 80525, a small business concern.

I hereby declare that the above-identified small business concern qualifies as a small business concern as defined in 13 CFR 121.3-18, and reproduced in 37 CFR 1.9(d), for purposes of paying reduced fees under section 41(a) and (b) of Title 35, United States Code, in that the number of employees of the concern, including those of its affiliates, does not exceed 500 persons. For purposes of this statement, (1) the number of employees of the business concern is the average over the previous fiscal year of the concern of the persons employed on a full-time, part-time or temporary basis during each of the pay periods of the fiscal year, and (2) concerns are affiliates of each other when either, directly or indirectly, one concern controls or has the power to control the other, or a third party or parties controls or has the power to control both.

I hereby declare that rights under contract or law have been conveyed to and remain with the small business concern identified above with regard to the invention, entitled "TICK OCTOPAMINE RECEPTOR NUCLEIC ACID MOLECULES, PROTEINS AND USES THEREOF," and identified as Attorney File No. P-FC-10, described in the specification filed herewith.

If the rights held by the above-identified small business concern are not exclusive, each individual, concern or organization having rights to the invention is listed below* and no rights to the invention are held by any person, other than the inventor, who could not qualify as a small business concern under 37 CFR 1.9(c) or by any concern which would not qualify as a small business concern under 37 CFR 1.9(d) or a nonprofit organization under 37 CFR 1.9(e).

*NOTE: Separate verified statements are required from each named person, concern or organization having rights to the invention averring to their status as small entities. (37 CFR 1.27)

NAME _____
 ADDRESS _____
☐ INDIVIDUAL ☐ SMALL BUSINESS CONCERN ☐ NONPROFIT
 ORGANIZATION

NAME _____
 ADDRESS _____
☐ INDIVIDUAL ☐ SMALL BUSINESS CONCERN ☐ NONPROFIT
 ORGANIZATION

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28(b))

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

Dated: November 15, 2002

By: Carol Talkington Verser
Carol Talkington Verser, Ph.D.
Executive Vice President
Intellectual Property and Business Development
Heska Corporation
1613 Prospect Parkway
Fort Collins, Colorado 80525

P-FC-10.ST25.txt
SEQUENCE LISTING

<110> Brandt, Kevin S.

<120> TICK OCTOPAMINE RECEPTOR NUCLEIC ACID MOLECULES, PROTEINS AND USES T
HEREOF

<130> P-FC-10

<140> not yet assigned

<141> 2002-11-15

<160> 21

<170> PatentIn version 3.1

<210> 1

<211> 102

<212> DNA

<213> Rhipicephalus sanguineus

<220>

<221> CDS

<222> (1) .. (102)

<223>

<400> 1
gcc atc att gtg ggc atc ttc atc gtg tgc tgg att aca gtg cat acc 48
Ala Ile Ile Val Gly Ile Phe Ile Val Cys Trp Ile Thr Val His Thr
1 5 10 15

ttt aga ttc ttt aag caa aca ctt ggc tac tgc aac tcg gcc ata aac
 Phe Arg Phe Phe Lys Gln Thr Leu Gly Tyr Cys Asn Ser Ala Ile Asn

20 25 30

96

ccc aga 102
Pro Arg

```
<210> 2
<211> 34
<212> PRT
<213> Rhipicephalus sanguineus
```

<400> 2

Ala Ile Ile Val Gly Ile Phe Ile Val Cys Trp Ile Thr Val His Thr
1 5 10 15

Phe Arg Phe Phe Lys Gln Thr Leu Gly Tyr Cys Asn Ser Ala Ile Asn
20 25 30

Pro Arg

P-FC-10.ST25.txt

<210> 3
<211> 102
<212> DNA
<213> Rhipicephalus sanguineus

<400> 3
tctgggggttt atggccgagt tgcagtagcc aagtgtttgc ttaaagaatc taaagggtatg 60
cactgtaatc cagcacacga tgaagatgcc cacaatgatg gc 102

<210> 4
<211> 499
<212> DNA
<213> Rhipicephalus sanguineus

<220>
<221> CDS
<222> (3)..(278)
<223>

<220>
<221> misc_feature
<222> (8)..(8)
<223> n = unknown

<400> 4
tg tct tan gag tca tca ccg tgg cat tcg ccc ttg gtg cgt gca ttc 47
Ser Xaa Glu Ser Ser Pro Trp His Ser Pro Leu Val Arg Ala Phe
1 5 10 15

tgc gag cac tgc atc ccc aac ctg ctg ttc tcg gtc ttc ttc tgg ctc 95
Cys Glu His Cys Ile Pro Asn, Leu Leu Phe Ser Val Phe Phe Trp Leu
20 25 30

gga tac tgc aac tcg gcc atc aac ccg ctc atc tac gtg ctt gtc agc 143
Gly Tyr Cys Asn Ser Ala Ile Asn Pro Leu Ile Tyr Val Leu Val Ser
35 40 45

aag gac ttt cgg ctg gcc ttc aag cgc atc ctg tgt cgc tgc cgc ctc 191
Lys Asp Phe Arg Leu Ala Phe Lys Arg Ile Leu Cys Arg Cys Arg Leu
50 55 60

aaa gaa gga ggc gtc tcg tca ctc atc aaa cag atc cac atg ctc acc 239
Lys Glu Gly Gly Val Ser Ser Leu Ile Lys Gln Ile His Met Leu Thr
65 70 75

gta ctt gac gac gca ccc ccg gac aac gcc gag tcg ccc tagaatcctg 288
Val Leu Asp Asp Ala Pro Pro Asp Asn Ala Glu Ser Pro
80 85 90

gtcacagcct tctcctgccc gcgctatcgc ggcattctca tggggcgcca ctgcttcttt 348

gcacctcacc cgctcactgc accgcgtgtc tctgcogtag tgacattgtc ggtgtccatc 408

tcacggctgt aatgtctcct tcctcaccca cgtgaatcac cactatagcc acagagcaaa 468

P-FC-10.ST25.txt

499

cgTgcCagta ccaagagcgc ttctgccgca c

<210> 5
 <211> 92
 <212> PRT
 <213> Rhipicephalus sanguineus
 <220>
 <221> misc_feature
 <222> (2)..(2)
 <223> The 'Xaa' at location 2 stands for Tyr
 <400> 5

Ser Xaa Glu Ser Ser Pro Trp His Ser Pro Leu Val Arg Ala Phe Cys
 1 5 10 15

Glu His Cys Ile Pro Asn Leu Leu Phe Ser Val Phe Phe Trp Leu Gly
 20 25 30

Tyr Cys Asn Ser Ala Ile Asn Pro Leu Ile Tyr Val Leu Val Ser Lys
 35 40 45

Asp Phe Arg Leu Ala Phe Lys Arg Ile Leu Cys Arg Cys Arg Leu Lys
 50 55 60

Glu Gly Gly Val Ser Ser Leu Ile Lys Gln Ile His Met Leu Thr Val
 65 70 75 80

Leu Asp Asp Ala Pro Pro Asp Asn Ala Glu Ser Pro
 85 90

<210> 6
 <211> 499
 <212> DNA
 <213> Rhipicephalus sanguineus

<220>
 <221> misc_feature
 <222> (492)..(492)
 <223> n = unknown

<400> 6
 gtgcggcaga agcgcctcttg gtactggcac gtttgctctg tggctatagt ggtgattcac 60
 gtgggtgagg aaggagacat tacagccgtg agatggacac cgacaatgtc actacggcag 120
 agacacgcgg tgcagtgagc ggggtgaggtg caaagaagca gtggcgcccc atgagaatgc 180
 cgcgatagcg cgggcaggag aaggctgtga ccaggattct agggcgactc ggcgttgtcc 240

P-FC-10.ST25.txt

```

gggggtgcgt cgtcaagtac ggtgagcatg tggatctgtt tgatgagtga cgagacgcct      300
ccttctttga ggccggcagcg acacaggatg cgcttgaagg ccagccgaaa gtccttgctg      360
acaagcacgt agatgagcgg gttgatggcc gagttgcagt atccgagcca gaagaagacc      420
gagaacagca ggttggggat gcagtgctcg cagaatgcac gcaccaaggg cgaatgccac      480
ggtgatgact cntaagaca                                                    499

```

```

<210> 7
<211> 286
<212> DNA
<213> Rhipicephalus sanguineus

```

```

<220>
<221> CDS
<222> (1)..(285)
<223>

```

```

<400> 7
cgg ccc ggt agc aac atg aac gag acg tgc ctg tcc cgc gtg ccg cca      48
Arg Pro Gly Ser Asn Met Asn Glu Thr Cys Leu Ser Arg Val Pro Pro
1 5 10 15

gag aag ctc cat gag ccg gtc act gtg gcc ctg ttc ttc gta ctg ggc      96
Glu Lys Leu His Glu Pro Val Thr Val Ala Leu Phe Phe Val Leu Gly
20 25 30

tcc atc aat gga ctc gtc atc ttc ggt aac ctg ctg gtc att atc gcc      144
Ser Ile Asn Gly Leu Val Ile Phe Gly Asn Leu Leu Val Ile Ile Ala
35 40 45

gtg ctg gcc tca aca aag ctg cgc acg gtc acc aac tac ttc gtg gtg      192
Val Leu Ala Ser Thr Lys Leu Arg Thr Val Thr Asn Tyr Phe Val Val
50 55 60

tcc ttg gct gtg gcc gac ctc tcg gtt ggg ctc acc gtg ttg cca tac      240
Ser Leu Ala Val Ala Asp Leu Ser Val Gly Leu Thr Val Leu Pro Tyr
65 70 75 80

tca att gtg ttg gag gtg ctc gag gtg tgg ctc ttc ggc caa cct g      286
Ser Ile Val Leu Glu Val Leu Glu Val Trp Leu Phe Gly Gln Pro
85 90 95

```

```

<210> 8
<211> 95
<212> PRT
<213> Rhipicephalus sanguineus

```

```

<400> 8
Arg Pro Gly Ser Asn Met Asn Glu Thr Cys Leu Ser Arg Val Pro Pro
1 5 10 15

```

P-FC-10.ST25.txt

Glu Lys Leu His Glu Pro Val Thr Val Ala Leu Phe Phe Val Leu Gly
 20 25 30

Ser Ile Asn Gly Leu Val Ile Phe Gly Asn Leu Leu Val Ile Ile Ala
 35 40 45

Val Leu Ala Ser Thr Lys Leu Arg Thr Val Thr Asn Tyr Phe Val Val
 50 55 60

Ser Leu Ala Val Ala Asp Leu Ser Val Gly Leu Thr Val Leu Pro Tyr
 65 70 75 80

Ser Ile Val Leu Glu Val Leu Glu Val Trp Leu Phe Gly Gln Pro
 85 90 95

<210> 9
 <211> 286
 <212> DNA
 <213> Rhipicephalus sanguineus

<400> 9
 cagggttgccc gaagagccac acctcgagca cctccaacac aattgagtat ggcaacacgg 60
 tgagcccaac cgagaggctcg gccacagcca aggacaccac gaagtagttg gtgaccgtgc 120
 gcagctttgt tgaggccagc acggcgataa tgaccagcag gttaccgaag atgacgagtc 180
 cattgatgga gcccagtacg aagaacaggg ccacagtgac cggctcatgg agcttctctg 240
 gcggcacgcg ggacaggcac gtctcgttca tgttgctacc gggccg 286

<210> 10
 <211> 1443
 <212> DNA
 <213> Rhipicephalus sanguineus

<220>
 <221> CDS
 <222> (1)..(1440)
 <223>

<400> 10
 atg aac gag acg tgc ctg tcc cgc gtg ccg cca gag aag ctc cat gag 48
 Met Asn Glu Thr Cys Leu Ser Arg Val Pro Pro Glu Lys Leu His Glu
 1 5 10 15
 ccg gtc act gtg gcc ctg ttc ttc gta ctg ggc tcc atc aat gga ctc 96
 Pro Val Thr Val Ala Leu Phe Phe Val Leu Gly Ser Ile Asn Gly Leu
 20 25 30
 gtc atc ttc ggt aac ctg ctg gtc att atc gcc gtg ctg gcc tca aca 144

P-FC-10.ST25.txt

Val	Ile	Phe	Gly	Asn	Leu	Leu	Val	Ile	Ile	Ala	Val	Leu	Ala	Ser	Thr		
	35						40					45					
aag	ctg	cgc	acg	gtc	acc	aac	tac	ttc	gtg	gtg	tcc	ttg	gct	gtg	gcc		192
Lys	Leu	Arg	Thr	Val	Thr	Asn	Tyr	Phe	Val	Val	Ser	Leu	Ala	Val	Ala		
	50					55					60						
gac	ctc	tcg	gtt	ggg	ctc	acc	gtg	ttg	cca	tac	tca	att	gtg	ttg	gag		240
Asp	Leu	Ser	Val	Gly	Leu	Thr	Val	Leu	Pro	Tyr	Ser	Ile	Val	Leu	Glu		
	65				70					75					80		
gtg	ctc	gag	gtg	tgg	atc	ttc	ggc	cac	acc	tgg	tgc	cag	ata	tgg	cta		288
Val	Leu	Glu	Val	Trp	Ile	Phe	Gly	His	Thr	Trp	Cys	Gln	Ile	Trp	Leu		
				85					90					95			
gct	gtg	gac	gta	tgg	ctc	tgc	aca	tca	tcc	atc	ctg	aat	ctc	tgc	gcc		336
Ala	Val	Asp	Val	Trp	Leu	Cys	Thr	Ser	Ser	Ile	Leu	Asn	Leu	Cys	Ala		
			100					105					110				
atc	agc	gtg	gac	cgc	tac	ttg	gcc	atc	acg	cgg	ccg	gtt	cgc	tac	cgg		384
Ile	Ser	Val	Asp	Arg	Tyr	Leu	Ala	Ile	Thr	Arg	Pro	Val	Arg	Tyr	Arg		
		115					120					125					
agc	ctc	atg	tcg	tcg	cgc	cgc	gcc	aag	ttg	ctc	atc	gtg	gcg	gtg	tgg		432
Ser	Leu	Met	Ser	Ser	Arg	Arg	Ala	Lys	Leu	Leu	Ile	Val	Ala	Val	Trp		
	130					135					140						
gtg	atc	gcc	ttt	gtc	atc	tgc	ttc	ccg	ccg	ctc	gtt	gga	tgg	aac	gac		480
Val	Ile	Ala	Phe	Val	Ile	Cys	Phe	Pro	Pro	Leu	Val	Gly	Trp	Asn	Asp		
	145				150					155					160		
ggc	ggg	tct	caa	aac	agc	gtg	cct	tac	cac	ggg	tcg	aac	gag	acc	ttg		528
Gly	Gly	Ser	Gln	Asn	Ser	Val	Pro	Tyr	His	Gly	Ser	Asn	Glu	Thr	Leu		
			165					170					175				
cac	aat	tcg	agc	atc	gct	gcc	gat	ggc	ccg	ttg	ccg	ctc	tgc	aag	tcc		576
His	Asn	Ser	Ser	Ile	Ala	Ala	Asp	Gly	Pro	Leu	Pro	Leu	Cys	Lys	Ser		
			180					185					190				
gca	caa	tgt	gtg	ctg	ata	aac	aac	aag	ggc	tac	gtc	atc	tac	tcg	gct		624
Ala	Gln	Cys	Val	Leu	Ile	Asn	Asn	Lys	Gly	Tyr	Val	Ile	Tyr	Ser	Ala		
		195					200					205					
ctg	ggc	tct	ttc	tac	ata	ccg	atg	ctg	ttc	atg	ctg	ttt	ttc	aac	tac		672
Leu	Gly	Ser	Phe	Tyr	Ile	Pro	Met	Leu	Phe	Met	Leu	Phe	Phe	Asn	Tyr		
	210					215					220						
cgc	ata	tac	cgc	gca	gcc	atc	cag	act	ggg	cgc	gcc	ctt	gaa	cga	ggt		720
Arg	Ile	Tyr	Arg	Ala	Ala	Ile	Gln	Thr	Gly	Arg	Ala	Leu	Glu	Arg	Gly		
	225				230				235						240		
ttc	ata	acc	aca	aag	tca	gga	aag	atc	aag	gga	cgc	aca	acg	gac	cag		768
Phe	Ile	Thr	Thr	Lys	Ser	Gly	Lys	Ile	Lys	Gly	Arg	Thr	Thr	Asp	Gln		
				245					250					255			
agg	ctc	acg	cta	cgc	gtc	cac	cgc	ggt	aac	gat	tcc	gct	atg	aac	gcc		816
Arg	Leu	Thr	Leu	Arg	Val	His	Arg	Gly	Asn	Asp	Ser	Ala	Met	Asn	Ala		
				260				265					270				

P-FC-10.ST25.txt

aag cga ggt agc gag cac ctc ggt gct gag acc tgc atc gat ggc atc	864
Lys Arg Gly Ser Glu His Leu Gly Ala Glu Thr Cys Ile Asp Gly Ile	
275 280 285	
gtc acc ggt cgt cgc cgg ccc gga ctd aag aag tgc cgc gac gag ccg	912
Val Thr Gly Arg Arg Arg Pro Gly Leu Lys Lys Ser Arg Asp Glu Pro	
290 295 300	
tca gcc agc gcc cgg tgc tca gcc agc aag acg cgt cag caa agt gac	960
Ser Ala Ser Ala Arg Ser Ser Ala Ser Lys Thr Arg Gln Gln Ser Asp	
305 310 315 320	
cag cgg acc acg cgc tgc tgc cgc ccc tgc ttc aag tgc aac agg ggc	1008
Gln Arg Thr Thr Arg Ser Ala Pro Pro Ser Phe Lys Ser Asn Arg Gly	
325 330 335	
agc gcc cgc aac agt gga cgc aac ggc act tcc acg tct agc ggc ggc	1056
Ser Ala Arg Asn Ser Gly Arg Asn Gly Thr Ser Thr Ser Ser Gly Gly	
340 345 350	
ggc aag ggc tgc cgt tgc agc aaa cgc agt caa cgg tgg cag gcc aag	1104
Gly Lys Gly Ser Arg Ser Lys Arg Ser Gln Arg Trp Gln Ala Lys	
355 360 365	
cga ttc cgc aca gag gcc aag gcc acc aag acc gtg ggc acc atc gtg	1152
Arg Phe Arg Thr Glu Ala Lys Ala Thr Lys Thr Val Gly Thr Ile Val	
370 375 380	
ggg gcc ttt ata tgc tgc tgg ctg ccc ttc ttc aca gtg tac ctg gtg	1200
Gly Gly Phe Ile Cys Cys Trp Leu Pro Phe Phe Thr Val Tyr Leu Val	
385 390 395 400	
cgt gca ttc tgc gag cac tgc atc ccc aac ctg ctg ttc tgc gtc ttc	1248
Arg Ala Phe Cys Glu His Cys Ile Pro Asn Leu Leu Phe Ser Val Phe	
405 410 415	
ttc tgg ctc gga tac tgc aac tgc gcc atc aac cgc ctc atc tac gtg	1296
Phe Trp Leu Gly Tyr Cys Asn Ser Ala Ile Asn Pro Leu Ile Tyr Val	
420 425 430	
ctt gtc agc aag gac ttt cgg ctg gcc ttc aag cgc atc ctg tgt cgc	1344
Leu Val Ser Lys Asp Phe Arg Leu Ala Phe Lys Arg Ile Leu Cys Arg	
435 440 445	
tgc cgc ctc aaa gaa gga ggc gtc tgc tca ctc atc aaa cag atc cac	1392
Cys Arg Leu Lys Glu Gly Gly Val Ser Ser Leu Ile Lys Gln Ile His	
450 455 460	
atg ctc acc gta ctt gac gac gca ccc cgc gac aac gcc gag tgc ccc	1440
Met Leu Thr Val Leu Asp Asp Ala Pro Pro Asp Asn Ala Glu Ser Pro	
465 470 475 480	
tag	1443

<210> 11
 <211> 480
 <212> PRT
 <213> Rhipicephalus sanguineus

P-FC-10.ST25.txt

<400> 11

```

Met Asn Glu Thr Cys Leu Ser Arg Val Pro Pro Glu Lys Leu His Glu
1          5          10          15

Pro Val Thr Val Ala Leu Phe Phe Val Leu Gly Ser Ile Asn Gly Leu
20          25          30

Val Ile Phe Gly Asn Leu Leu Val Ile Ile Ala Val Leu Ala Ser Thr
35          40          45

Lys Leu Arg Thr Val Thr Asn Tyr Phe Val Val Ser Leu Ala Val Ala
50          55          60

Asp Leu Ser Val Gly Leu Thr Val Leu Pro Tyr Ser Ile Val Leu Glu
65          70          75          80

Val Leu Glu Val Trp Ile Phe Gly His Thr Trp Cys Gln Ile Trp Leu
85          90          95

Ala Val Asp Val Trp Leu Cys Thr Ser Ser Ile Leu Asn Leu Cys Ala
100         105         110

Ile Ser Val Asp Arg Tyr Leu Ala Ile Thr Arg Pro Val Arg Tyr Arg
115         120         125

Ser Leu Met Ser Ser Arg Arg Ala Lys Leu Leu Ile Val Ala Val Trp
130         135         140

Val Ile Ala Phe Val Ile Cys Phe Pro Pro Leu Val Gly Trp Asn Asp
145         150         155         160

Gly Gly Ser Gln Asn Ser Val Pro Tyr His Gly Ser Asn Glu Thr Leu
165         170         175

His Asn Ser Ser Ile Ala Ala Asp Gly Pro Leu Pro Leu Cys Lys Ser
180         185         190

Ala Gln Cys Val Leu Ile Asn Asn Lys Gly Tyr Val Ile Tyr Ser Ala
195         200         205

Leu Gly Ser Phe Tyr Ile Pro Met Leu Phe Met Leu Phe Phe Asn Tyr
210         215         220

```

P-FC-10.ST25.txt

Arg Ile Tyr Arg Ala Ala Ile Gln Thr Gly Arg Ala Leu Glu Arg Gly
225 230 235 240

Phe Ile Thr Thr Lys Ser Gly Lys Ile Lys Gly Arg Thr Thr Asp Gln
245 250 255

Arg Leu Thr Leu Arg Val His Arg Gly Asn Asp Ser Ala Met Asn Ala
260 265 270

Lys Arg Gly Ser Glu His Leu Gly Ala Glu Thr Cys Ile Asp Gly Ile
275 280 285

Val Thr Gly Arg Arg Arg Pro Gly Leu Lys Lys Ser Arg Asp Glu Pro
290 295 300

Ser Ala Ser Ala Arg Ser Ser Ala Ser Lys Thr Arg Gln Gln Ser Asp
305 310 315 320

Gln Arg Thr Thr Arg Ser Ala Pro Pro Ser Phe Lys Ser Asn Arg Gly
325 330 335

Ser Ala Arg Asn Ser Gly Arg Asn Gly Thr Ser Thr Ser Ser Gly Gly
340 345 350

Gly Lys Gly Ser Arg Ser Ser Lys Arg Ser Gln Arg Trp Gln Ala Lys
355 360 365

Arg Phe Arg Thr Glu Ala Lys Ala Thr Lys Thr Val Gly Thr Ile Val
370 375 380

Gly Gly Phe Ile Cys Cys Trp Leu Pro Phe Phe Thr Val Tyr Leu Val
385 390 395 400

Arg Ala Phe Cys Glu His Cys Ile Pro Asn Leu Leu Phe Ser Val Phe
405 410 415

Phe Trp Leu Gly Tyr Cys Asn Ser Ala Ile Asn Pro Leu Ile Tyr Val
420 425 430

Leu Val Ser Lys Asp Phe Arg Leu Ala Phe Lys Arg Ile Leu Cys Arg
435 440 445

Cys Arg Leu Lys Glu Gly Gly Val Ser Ser Leu Ile Lys Gln Ile His
450 455 460

P-FC-10.ST25.txt

Met Leu Thr Val Leu Asp Asp Ala Pro Pro Asp Asn Ala Glu Ser Pro
 465 470 475 480

<210> 12
 <211> 1443
 <212> DNA
 <213> *Rhipicephalus sanguineus*

<400> 12
 ctagggcgac tcggcggtgt ccgggggtgc gtcgtcaagt acggtgagca tgtggatctg 60
 tttgatgagt gacgagacgc ctcttctttt gaggcggcag cgacacagga tgcgcttgaa 120
 ggccagccga aagtccttgc tgacaagcac gtagatgagc gggttgatgg ccgagttgca 180
 gtatccgagc cagaagaaga ccgagaacag caggttgggg atgcagtgtc cgcagaatgc 240
 acgcaccagg tacactgtga agaagggcag ccagcagcat ataaagccac ccacgatggt 300
 gccacggtc ttggtggcct tggcctctgt gcggaatcgc ttggcctgcc accgttgact 360
 gcgtttgctc gaacgcgagc ccttgccgcc gccgctagac gtggaagtgc cgttgcgctc 420
 actgttgccg gcgctgcccc tgttcgactt gaacgagggc ggccgagcgc gcgtgggtccg 480
 ctggtcactt tgctgacgcg tcttgctggc tgacgaccgg gcgctggctg acggctcgtc 540
 gcgcgacttc ttgagtcggg gccggcgacg accggtgacg atgccatcga tgcaggtctc 600
 agcaccgagg tgctcgctac ctgccttggc gttcatagcg gaatcggtac cgcgggtggac 660
 gcgtagcgtg agcctctggg ccgttggtgcg tcccttgatc tttcctgact ttgtgggtat 720
 gaaacctcgt tcaagggcgc gccagtcctg gatggctgcg cggatatatgc ggtagttgaa 780
 aaacagcatg aacagcatcg gtatgtagaa agagcccaga gccgagtaga tgacgtagcc 840
 cttgttgttt atcagcacac attgtgcgga cttgcagagc ggcaacgggc catcggcagc 900
 gatgctcgaa ttgtgcaagg tctcgttcga ccggtggtaa ggcacgctgt tttgagacct 960
 gccgtcgctc catccaacga gcggcgggaa gcagatgaca aaggcgatca cccacaccgc 1020
 cacgatgagc aacttggcgc ggccgacga catgaggctc cggtagcgaa ccggccgcgt 1080
 gatggccaag tagcgggtcca cgctgatggc gcagagattc aggatggatg atgtgcagag 1140
 ccatacgctc acagctagcc atatctggca ccagggtgtg ccgaagatcc acacctcgag 1200
 caacctcaac acaattgagt atggcaacac ggtgagccca accgagaggt cggccacagc 1260
 caaggacacc acgaagtagt tggtgaccgt gcgcagcttt gttgaggcca gcacggcgat 1320
 aatgaccagc aggttaccga agatgacgag tccattgatg gagcccagta cgaagaacag 1380
 gccacagtg accggctcat ggagcttctc tggcggcacg cgggacaggc acgtctcgtt 1440

P-FC-10.ST25.txt

1443

cat

<210> 13
 <211> 33
 <212> DNA
 <213> Artificial sequence

<220>
 <223> Synthetic Primer

<400> 13
 gccatcatyg tgggcrkstt catcktbgtgc tgg

33

<210> 14
 <211> 33
 <212> DNA
 <213> Artificial sequence

<220>
 <223> Synthetic Primer

<400> 14
 gatcatsggr ttwayggcsg agttgcagta gcc

33

<210> 15
 <211> 23
 <212> DNA
 <213> Artificial sequence

<220>
 <223> Synthetic Primer

<400> 15
 gctggctgcc attcttcacc gtg

23

<210> 16
 <211> 27
 <212> DNA
 <213> Artificial sequence

<220>
 <223> Synthetic Primer

<400> 16
 ccataccta acgactcact atagggc

27

<210> 17
 <211> 24
 <212> DNA
 <213> Artificial sequence

<220>
 <223> Synthetic Primer

P-FC-10.ST25.txt

<400> 17
ggtgcgtgca ttctgcgagc actg 24

<210> 18
<211> 24
<212> DNA
<213> Artificial sequence

<220>
<223> Synthetic Primer

<400> 18
agaagaccga gaacagcagg ttgg 24

<210> 19
<211> 29
<212> DNA
<213> Artificial sequence

<220>
<223> Synthetic Primer

<400> 19
tggcaccagg tgtggccgaa gagccacac 29

<210> 20
<211> 24
<212> DNA
<213> Artificial sequence

<220>
<223> Synthetic Primer

<400> 20
atgaacgaga cgtgcctgtc ccgc 24

<210> 21
<211> 24
<212> DNA
<213> Artificial sequence

<220>
<223> Synthetic Primer

<400> 21
ctagggcgac gcggcgttgt ccgg 24